EU COST Action FA0701 Summit meeting

Arthropod Symbiosis: from fundamental research to pest and disease management

St. Pierre d' Oléron, France

12-13 of June, 2012

Organizing Committee

Myriam BADAWI, Nicolas BECH, Sophie BELTRAN, Joanne BERTAUX, Didier BOUCHON, Christine BRAQUART-VARNIER, Yves CAUBET, Richard CORDAUX, Catherine DEBENEST, Carine DELAUNAY, Jessica DITTMER, Sandrine GENIEZ, Lise GENTY, Clément GILBERT, Isabelle GIRAUD, Frédéric GRANDJEAN, Pierre GREVE, Monique JOHNSON, Alexandra LAFITTE, Tiffany LAVERRÉ, Winka Le CLEC'H, Jérôme LESOBRE, Isabelle MARCADÉ, Christelle MIREBEAU, Bouziane MOUMEN, Elmina MOTTIN, Maryline RAIMOND, Roland RAIMOND, Freddie-Jeanne RICHARD, Mathieu SICARD, Catherine SOUTY-GROSSET

> from UMR CNRS 7267 Ecologie et Biologie des Interactions, équipe Ecologie Evolution Symbiose POITIERS

With the participation of UMR CNRS 5558 Laboratoire de Biométrie et Biologie Évolutive LYON, UMR5557- Ecologie Microbienne LYON, UMR INRA/INSA Biologie Fonctionnelle Insectes et Interactions LYON, Institut des Sciences de l'Evolution MONTPELLIER

Scientific Committee

Didier BOUCHON (University of Poitiers, FRANCE), Kostas BOURTZIS (University of Western Greece, GREECE), Sylvain CHARLAT (University of Lyon, FRANCE), Richard CORDAUX (University of Poitiers, FRANCE), Stephen DOBSON (University of Kentucky, USA), Olivier DURON (University of Montpellier, FRANCE), Takema FUKATSU (National Institute of Adavanced Industrial Science and Technology, JAPAN), Pierre GREVE (University of Poitiers, FRANCE), Abdelaziz HEDDI (INSA of Lyon, FRANCE), Patrick MAVINGUI (University of Lyon, FRANCE), Barton SLATKO (New England Biolabs, USA), Einat ZCHORI-FEIN (Agricultural Research Organization, ISRAEL)

> This book was designed by Christelle Mirebeau, EBI, UMR CNRS 7267, Poitiers Imprimerie Copy-Media - Mérignac

June 11th : Arrival day

June 12th

8:30-9:00 Kostas Bourtzis & Einat Zchori-Fein (Chair & Vice Chair of the COST Action): Welcome address and summary of COST Action

Honoring Lynn Margulis

- 9:00-9:40 Antonio Lazcano: Symbiosis and evolution: on the origin of cells
- 9:40-10:00 **Claudio Bandi:** Journeys with a free mind: life, symbiogenesis and our personal memories of Lynn Margulis

10:00-10:25 : coffee break

WP1 Arthropod Symbiont Diversity

- 10:25-10:40 WG leaders (Daffonchio & Horn): Overview of WG1
- 10:40-11:10 **Takema Fukatsu:** Novel experimental model systems for insect symbiosis studies
- 11:10-11:30 Koos Boomsma: On ant symbiotic associations
- 11:30-11:50 **Patrick Mavingui:** Symbiotic diversity in mosquito vectors from Madagascar and Reunion islands, Indian Ocean
- 11:50-12:10 **Olle Terenius:** Thorsellia anophelis and Aedes aegypti symbionts
- 12:10-12:30 **Rok Kostanjsek:** Recent advances and future perspectives of *Rhabochlamydia porcellionis* research
- 12:30-12:50 Hans Breeuwer: Symbionts in mites

13:00-14:00 : Lunch

WP2 Arthropod-Symbiont Metagenomes

- 14:00-14:15 WG leaders (Anderson & Lattore): Overview of WG2
- 14:15-14:45 Serap Aksoy: Tsetse symbiosis with genus Wigglesworthia
- 14:45-15:05 Lisa Klasson: Comparative genomics of insect endosymbionts
- 15:05-15:25 **Matthias Horn:** *'Candidatus Cardinium hertigii'* a reproductive manipulator with a role in host nutrition
- 15:25-15:45 Alistair Darby: Arsenophonus genomes

15:45-16:00 : coffee break

- 16:00-16:20 Fabrice Vavre: The story of communities affecting communities
- 16:20-16:40 **Sandy Macdonald:** Nitrogen recycling in the aphid/*Buchnera* symbiosis: the role of the host cell

WP3 Host-Symbiont Interactions

- 16:40-16:55 WG leaders (Gross, Hurst & Vavre): Overview of WG3
- 16:55-17:25 Jan van den Abbeele: Sodalis glossinidius, a symbiotic Trojan horse against a tsetse fly transmitted parasite - the trypanosome
- 17:25-17:55 Abdelaziz Heddi: Immunity and symbiosis in weevils

18:00-20:00 : Final MC meeting

20:00 :Dinner

June 13th

- 8:30-8:50 **Amparo Lattore:** From insect pathogen to obligate symbionts: what's in between?
- 8:50-9:10 **Carolin Ratzka:** Immune response of the ant *Camponotus floridanus* against pathogens and its obligate mutualistic endosymbiont
- 9:10-9:30 Christoph Vorburger: Symbionts mediate coevolution between aphid hosts and parasitoids
- 9:30-9:50 **Greg Hurst:** Disease epidemiology in arthropods is altered by the presence of non-protective symbionts
- 9:50-10:10 **Mathieu Sicard:** Experimental evolution of virulence in a symbiosis: from vertical to horizontal transmission

10:10-10:30 : coffee break

WP4 Symbiont-Based Control Strategies

- 10:30-10:45 WG4 leaders (Bandi & Sinkins): Overview of WG4
- 10:45-11:15 Horacio Frydman: Wolbachia as a tool for stem cell biology studies
- 11:15-11:35 **Daniele Daffonchio:** Is the "multicoloured" microbiome associated with insect vectors of plant pathogens a potential resource for disease control?
- 11:35-11:55 **Guido Favia:** Mosquitoes-symbionts interactions: from complex relationships to biotechnological applications
- 11:55-12:15 Ameur Cherif: Symbiotic control of honeybee pathogens
- 12:15-12:35 **Boaz Yuval:** Tephritid symbionts and their applied potential
- 12:35-12:55 **Tom Groot**: The potential of symbionts in improving biological control agents

13:00-14:10 : Lunch

WP5 Regulations

- 14:10-14:40 **Jaime Aguilera:** Regulatory aspects and safety considerations in the EU
- 14:40 15:00 Maria Alejandra Perotti: Mites, symbionts and commercial implications
- 15:00-15:20 **Henk Braig:** Is it mine? Ethical, regulatory and other considerations regarding the manipulation of the human microbiome or human symbionts

15:20-15:55 : coffee break

National projects of COST partners

- 15:55-16:10 Liga Jankevica: Research on arthropod symbiosis in Latvia
- 16:10-16:25 Sandra Breum Andersen: Dynamic Wolbachia prevalence in Acromyrmex leaf-cutting ants

ESRs presenting STSMs

- 16:25-16:40 STSM Coordinator (Feldhaar & Wilkinson): Overview of STSMs
- 16:40-16:55 **Renate Zindel:** The microbiome of the pest mite *Rhizoglyphus robini*: implications for the pest's status
- 16:55-17:10 **Davide Sassera:** The genome sequence of *Midichloria mitochondrii* and the origin of mitochondria
- 17:10-17:25 **Evangelos Doudoumis:** On tsetse flies-*Wolbachia* symbiotic associations
- 17:25-17:40 **Marta Moreira:** Detection of spiders microbial communities in Madeira Island
- 17:40-17:55 Anton Strunov: Effect of high temperature on pathogenic Wolbachia strain wMelPop in Drosophila melanogaster brain cells
- 17:55-18:10 Daniela Schneider: Manipulation of mating behavior
- 18:10-18:25 **Thijs Machtelinckx:** Effects of bacterial endosymbionts on the use of predatory bugs in biological pest control
- 18:30-20:00 Poster session

18:30-20:00 <u>Evaluation meeting</u> (Participants: Prof. Anna Pretova, Dr. Ioanna Stavridou, Prof. Kostas Bourtzis, Dr. Einat Zchori-Fein and External Evaluator)

20:00 : Dinner

June 14th : Departure day

Oral Presentations

EU COST Action FA0701 «Arthropod Symbiosis: from fundamental research to pest and disease management»: a synopsis of a four-year-long journey

Bourtzis K.^{1,2} and Zchori-Fein E.³

¹Department of Environmental and Natural Resources Management, University of Western Greece, 2 Seferi St, 30100 Agrinio, Greece; ² Biomedical Sciences Research Center Al. Fleming, 16672 Vari, Greece; ³Agricultural Research Organization, Entomology, Newe Ya'ar Research Center 30095 Ramat Yishay Israel - kbourtz@uoi.gr

During the last decades, there has been growing concern in the medical and agricultural communities about the many risks associated with chemical pest and disease control. Certain arthropods are devastating agricultural pests, others are exceptionally efficient vectors of plant, animal and human pathogens. Bacterial symbionts are present in many of these arthropod pests. Manipulation of the symbiotic relationships could provide novel and revolutionary biotechnological means for improving pest control for food production, public health and environment protection. The COST Action FA0701 "Arthropod Symbiosis: from fundamental research to pest and disease management" was funded by the EU with the goal of exploring and harnessing arthropod symbiotic associations, from the level of single genes through genomics to populations, towards the development and commercialization of symbiont-based control strategies (SCS) that are environmentally friendly and have the potential to replace chemical control methods. Major aspects of the progress made during the four years of our Action (26 March 2008 - 25 March 2012) towards achieving these goals will be presented.

	<hr/>
/	
1)
\vdash	
	1
\vdash	
L	
\vdash	
	1
\vdash $ -$	
1	
	A
1	
1	
\	/
\langle	
	/
	-

Symbiosis and evolution: on the origin of cells

Lazcano A.

Facultad de Ciencias, UNAM, Mexico - alar@ciencias.unam.mx

In 1967 Lynn Margulis published a now classical paper in the *Journal of Theoretical Biology* advocating the endosymbiotic origin of eukaryotic cells. While it can be argued that some of these ideas had been suggested since the late 19th century, it is equally true that most of them were largely speculative proposals part of the non Darwinian explanations prevalent during such period. In contrast, Lynn Margulis provided a testable hypothesis that made specific predictions in a broader context, and that placed the evolution of cells within a wider context. It is little appreciated that when Lynn Margulis' first published her proposal, the molecularization of life sciences was undergoing an explosive development. Much was gained, but often at the expense of other approaches including biochemistry, organismal biology, and an evolutionary perspective. Equally significant, neodarwinism was at its height, and it became difficult for many not only to accept the idea of endosymbiosis but also to acknowledge that microbes where not only pathogens, but also the ancestors of all extant life.

Lynn Margulis's first book is remarkable. It discusses not only prokaryotic evolution and the origin of nucleated cells, but also the deep interaction between the biosphere and the planet itself. This volume is a sample of impressive scholarship and intellectual versatility, in which she traced in authoritative detail the development of microbial life and the emergence of eukaryotes as the outcome of symbiosis. One of the one of the most notable aspects of the books its is multidisciplinary approach, linking biological evolution to the transformation of the Earth into a planet endowed with an oxygen atmosphere and the subsequent changes in the chemistry of the oceans and sediments.

Her views do not contradict Darwinian evolution by natural selection. Endosymbiosis represents a different form of genetic variation, and like conjugation and meiotic sex followed by fertilization, it is a combinatorial mode of evolutionary change. The statement that symbiosis is a Lamarckian event is certainly incorrect. Indeed, the acquisition of genes and genomes due to endosymbiosis is a random event, i.e., it is independent of the advantage it may confer on the organisms in their specific environment, and the biological traits acquired by endosymbiotic events have a previous history, which has been shaped by the sieve of natural selection. What Lynn Margulis did was not only to document symbiosis as a major source of biological variation and evolution, but also to change our views of the biosphere itself.

Keywords: endosymbiosis, Margulis, Darwinian evolution

Journeys with a free mind: life, symbiogenesis and our personal memories of Lynn Margulis

Bandi C.¹, Pajoro M.², Sassera D.¹ and Sacchi L.³

¹Università degli Studi di Milano; ²IZSLER, Sezione di Pavia; ¹Università degli Studi di Pavia - claudio.bandi@unimi.it

In the field of evolutionary biology, dominated by mathematical models and equations, aimed at describing the dynamics of micromutations within populations, the study of symbiosis remained neglected for years. It was useful to provide students with nice examples and curiosities, but was not regarded as a field capable of generating general concepts on evolution. In this framework, till about 20 years ago, Lynn Margulis' seminal papers and books on the symbiotic origin of eukaryotic cells seem to have been considered only in piecemeal fashion, limited to the origin of mitochondria and chloroplasts. Studies published in different fields in the last 20/30 years have however lead to a profound re-evaluation of symbiosis, and have convinced most biologists that symbiosis acted and still acts as a central force in evolution. Lynn Margulis is now part of the history of biology, for the contributions she gave to symbiosis and to the understanding of the origin of eukaryotic cells. But there are other aspects of the science and culture and of Lynn Margulis that are less known, and we will devote part of the seminar to these aspects. Lynn Margulis had a profound knowledge and passion for a variety of microorganisms, from prokaryotes to eukaryotic algae, and for a variety of aspects of organismal biology and ecology. From the details of a specific biological system, she had an unsurpassed capacity to derive general concepts, and to integrate the different tiles of the mosaic into a unique, coherent picture. It is thus not a case that she loved pointillist painters, like Seurat, Signac, Pelizza da Volpedo, that portrayed humans, animals, plants and their environment using collections of tiny points. The stories, or the 'points', that formed the evolving and living paint of Lynn Margulis were Mastotermes darwiniensis, Mixotricha paradoxa, Treponema pallidum, Pectinatella magnifica, and many others. We learned some of these stories during a trip by car with Lynn Margulis, from Milano to Pisa. She was going receive a Laurea ad Honorem at the University of Pisa, and she asked one of us to translate into Italian the talk she had prepared for the ceremony. The four hours of driving in the car, while translating a talk and explaining Italian pronunciation, had been very intense, and we had a unique, crazy and funny occasion for understanding a little piece of Lynn Margulis' view of life.

Keywords: symbiogenesis, eukaryotic cell, spirochetes, Midichloria, syphilis

Novel experimental model systems for insect symbiosis studies

Fukatsu T.

National Institute of Advanced Industrial Science and Technology (AIST), Tsukuba 305-8566, Japan - t-fukatsu@aist.go.jp

There are several well-established insect models for symbiosis studies, represented by aphid-Buchnera for obligate symbiotic association and Drosophila-Wolbachia for facultative symbiotic association, with both host and symbiont genomic data available. In addition, powerful molecular and genetic tools are available for the Drosophila system. Meanwhile, symbiont culturing and genetic manipulation are practically impossible in both the systems. In the Drosophila system, highly-developed symbiotic organs are absent. In the aphid system, even RNA interference does not work, which makes functional analyses quite difficult. To overcome these difficulties, we are recently developing several new insect models with the following features: (i) the host insects are easily maintainable and experimentally tractable in the laboratory, (ii) the symbiotic bacteria are easily cultivable on standard microbiological media, (iii) genetic transformation and gene-knockout are applicable to the symbiotic bacteria, (iv) the genetically manipulated symbiotic bacteria are easily re-introduced into the host insects, (v) the complete symbiont genomes are being determined, (vi) the RNA sequencing data are being accumulated for their well-developed symbiotic organs, and (vii) expression of the host insect genes can be efficiently suppressed by RNA interference. We present several remarkable achievements with these new insect models for understanding of molecular mechanisms underlying the insect-microbe associations.

Keywords : new insect models, symbiont culturing and genetic manipulation, symbiont genomics, host transcriptomics, RNA interference

Ant bacterial symbionts: how specific can they be and do they need to be?

Boomsma Jacobus J. (Koos), Andersen S.B. and Sapountzis P.

Centre for Social Evolution, Department of Biology, University of Copenhagen, Universitetsparken 15, 2100 Copenhagen - JJBoomsma@ bio.ku.dk

In recent years, many remarkable bacterial symbionts have been discovered to be associated with ants. They are usually related to diet deficiencies, particularly for ants that abandoned their original wasp-like predatory life-style to become dependent on homopteran secretions, but the protective association between fungus-growing ants and cuticular cultures of actinomycetes has also attracted considerable interest and debate. Evolutionary theory predicts that symbionts with the same host niche should end up being kept in monocultures, but selection in this direction will continue to be challenged by infections with competing strains in spite of expected countermeasures by hosts and resident symbionts. Recent results from 454 sequencing show that the cuticular ectosymbionts of Acromyrmex leaf-cutting ants are much more specific and monocultural than recent challenges of this idea have suggested. We hypothesize that the unusually constant and possibly partially deficient diet of leaf-cutting ants should have provided interesting additional niches for bacterial symbionts. The first findings confirming this idea will be presented and discussed in light of the title of this presentation.

Keywords: fungus-growing ants, symbiosis, actinomycetes, genome sequencing, diet

		<hr/>
/		
		1
		1
		1
		1
		1
		1
		1
		1
		1
		1
		1
		1
		1
		1
		1
		1
		1
		1
		1
		1
		1
		1
		1
		1
		1
		1
		1
		1
	~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	1
		1
\		/
$\backslash$		
		/

# Symbotic diversity in mosquito vectors from Madagascar and Reunion islands, Indian Ocean

Zouache K., Raharimalala F., Raveloson L., Tran-Van V., Failloux A-B., Ravelonandro P. and <u>Mavingui P.</u>

#### Ecologie Microbienne Lyon, UMR CNRS 5557, Université Lyon 1, 69622 VILLEURBANNE FRANCE - patrick.mavingui@univ-lyon1.fr

Symbiotic bacteria have a considerable impact on the behavior of many arthropods hosts. Indeed, it is well known that microbial communities associated with arthropods can contribute to host reproduction and survival, community interactions, protection against natural enemies and vectorial competence. However, such extended phenotypes were mostly shown in phytophagous insects, whereas research on hematophagous insects has been limited. For instance, the current knowledge of bacterial communities associated with mosquitoes is very limited and consequently their contribution to host behaviors is mostly unknown. As recent works have demonstrated that some Wolbachia strains can inhibit mosquito-borne pathogens in some circumstances, characterizing the microbiote of mosquitoes in their environment is a step forward in understanding the ecology and the multipartite interactions occurring in these vectors. In this research we surveyed the composition of bacterial communities associated with wild mosquitoes and explored whether the bacterial diversity can be related to host ecology and pathogen infection. To that purpose, we have chosen two insular ecosystems, Madagascar and Reunion islands, that are prone to invasion by mosquito vectors and where severe epidemics of dengue and chikungunya occurred recently involving high incidences in human populations.

Keywords: Aedes aegypti, Aedes albopictus, Microbial ecology, Symbiosis

### Thorsellia anophelis and Aedes aegypti symbionts

<u>Terenius O.</u>¹, Linda J.M.², Guy L.³, Eriksson-Gonzales K.⁴, Bussière L.⁵, Laugen A.T.¹, Titanji K.⁶ and Faye I.⁴

¹Department of Ecology, Swedish University of Agricultural Sciences (SLU), Uppsala, Sweden; ²Department of Chemistry, Royal Institute of Technology, Stockholm, Sweden; ³Department of Molecular Evolution, EBC, Uppsala University, Uppsala, Sweden; ⁴Department of Genetics, Microbiology and Toxicology, Stockholm University, Sweden; ⁵Institute of Biological and Environmental Sciences, University of Stirling, United Kingdom; ⁶Department of Microbiology and Immunology, Emory University School of Medicine, Atlanta, GA, USA - olle.terenius@slu.se

In our research we have focused on the possible co-adaptation of gut bacteria to their mosquito host. We have previously isolated a unique species of bacteria, *Thorsellia anophelis*, from Anopheles *gambiae* in Kenya. *T. anophelis* has also been found to dominate the gut flora of *An. gambiae* in two other locations in Kenya. Recent data show that *Thorsellia* species also exist in the major malaria vectors of South America and Asia, but so far it has not been found outside malaria mosquitoes or their breeding waters. We have genome sequenced the type isolate from Kenya discovering that it belongs to a new family of bacteria within gamma-proteobacteria.

We have also experimentally analyzed midgut bacteria growth in *Aedes aegypti*. To quantify the dynamics of midgut bacteria growth we experimentally infected *Ae. aegypti* with four different bacterial species isolated from five different host species. We show that the four bacterial species differ in dynamics over time and both the dynamics over time as well as the differences between the species depend on host sex. Our results can at least partly be explained by co-adaptation between host and bacteria and suggest that even closely related species of bacteria differ in their ability to remain in the mosquitoes.

Keywords : Thorsellia, Anopheles, Pantoea, Enterobacter, Aedes aegypti

# Recent advances and future perspectives of *Rhabochlamydia porcellionis* research

#### Kostanjsek R.

#### Department of Biology, Biotechnical Faculty, University of Ljubljana, Vecna pot 111, 1000 Ljubljana, Slovenia - rok.kostanjsek@bf.uni-lj.si

*Rhabdochlamydia porcellionis* is intracellular pathogen initially described in digestive glands or hepatopancreas of terrestrial isopod crustacean *Porcellio scaber*. Beside the later, the infections with the members of the family Rhabdochlamydiaceae are described in other arthropods, as well as in amoebe from water, soil and sponges. At the same time recent description of Rhabdochlamydia-like sequences in clinical isolates of vertebrates, including humans indicate their pathogenic potential in higher animals as well. In order to obtain additional data on their transmission, host range and interactions with host, it is essential to maintain the rhabdochlamydia in pure culture, which is, according to their intracellular nature, possible only with their cocultivation within cell lines.

In recent studies we combined the expertise of recognition and isolation of infected tissues in isopods, with co-cultivation of environmental chlamydiae in insect cell lines. Successfully infected cell lines were used to optimize the stringency of hybridization conditions by *R. porcellionis*-specific probe in FISH, develop the protocol for fixation and preservation of *R. porcellionis* morphology in cell lines for further ultrastructural analysis by electron microscopy, and to optimize a protocol for isolation of high molecular weight DNA from co-cultivated *Rhabdochlamydia*, required for its further genome sequencing analysis. Successful isolation of *R. porcellionis* in insect cell lines therefore provides important step towards understanding the biology of these intracellular bacteria, which will initially include comparative genomics and proteomics, as well as ultrastructural description of *R. porcellionis* developmental cycle, which is already in progress.

Keywords : Rhabdochlamydia porcellionis, cell lines, isolation, Porcellio scaber

### Symbionts in mites

#### Breeuwer J.A.J.

# Institute for Biodiversity and Ecosystem Dynamics, University of Amsterdam, The Netherlands - J.A.J.Breeuwer@uva.nl

Mites can be infected with bacteria that can alter the normal mode of reproduction. These bacteria are Wolbachia and Cardinium, but more may be found since only a small number of mite species have been studied. Also mite endosymbionts seem to be more difficult to detect by specific PCR, because they either occur at lower density or are slightly different from similar symbionts in insect hosts. I will discuss two examples of symbiont induced parthenogenesis in mites: the genus *Brevipalpus*, infected with *Cardninium*, and the genus Bryobia infected with Wolbachia. The genus Brevipalpus is characterized by a unique feature: they are the only known animal species with haploid females. This situation is caused by an infection with intracellular Cardinium bacteria. These bacteria feminize haploid males, thereby ensuring that they can be vertically transmitted to the next generation. An mtDNA phylogeny shows that Brevipalpus mites can be divided into a limited number of clades, representing the most abundant species. Three species are infected and feminized by Cardinium symbionts. In the fourth species the feminization appeared to be a genetic trait of the mite itself, since no Cardinium is present. Comparing mitochondrial and nuclear genetic markers showed that two species have reproduced strictly clonally for guite some time. In the other two species evidence was found for sexual reproduction in the relatively recent past. In the genus Bryobia, several sexual and parthenogenetic species have been found. Parthenogenesis is induced by Wolbachia, but double infections with Cardinium do occur. Also, double infections are found in sexual species that induce incompatibility. I will argue that asexuality originated multiple times within Bryobia. Wolbachia bacteria cause asexuality in at least two Bryobia species and may have infected different species independently. Although asexual reproduction is expected to result in low levels of genetic variation, we found a large number of clones in each species. Incongruencies between the mitochondrial phylogeny of the host and bacterial phylogeny, suggests that there may be occasional sexual reproduction or horizontal transfer and recombination of symbionts. Both suggestions will generate genetic variation.

### Tsetse symbiosis with genus Wigglesworthia

Brian W.¹, Wang J.¹, Brelsfoard C.^{1,2} and <u>Aksoy S.¹</u>

¹Yale University School of Public Health, 60 College St., 626 LEPH, New Haven, CT 06520 USA; ²Current address: Department of Entomology, University of Kentucky, S-225 Ag. Science Center North, Lexington, KY 40546 USA - serap.aksoy@yale.edu

Tsetse flies (*Glossina* spp.) are the sole vectors of *Trypanosoma* spp., the causative agents of sleeping sickness in humans (human African trypanosomosis, HAT) and of nagana (African animal trypanosomosis, AAT) in livestock. Although epidemics have significantly declined during the last years, still tens of millions of people in Africa continue to be at risk of contracting sleeping sickness. While control of disease in the mammalian host is difficult, reduction of tsetse populations is an effective approach for disease control. In this regards, tsetse's dependence on its microbiome provides novel opportunities for control.

Wigglesworthia is an obligate inherited symbiont of tsetse flies. Wigglesworthia resides a group cells (bacteriocytes) forming the midgut bacteriome organ. In addition, extracellular forms of the bacterium are present in the milk secretions of the accessary milk gland, which provides the route of transmission to tsetse's intrauterine larva. The genome sequence indicates retension of vitamin pathways, suggestions a nutritional role for this organism to supplement tsetse's vertebrate specific bloodmeal diet.

Our study has indicated that elimination of *Wigglesworthia* by antibiotic treatment coupled with dietary supplementation of yeast extract gives rise to female progeny that are infertile but that also lack immunity functions, particularly cellular immunity. Supplementation of tsetse's diet with antibiotics, yeast and *Wigglesworthia* extracts can rescue loss of immunity as detected by increased presence of hemocytes in sterile progeny. A tsetse protein, Peptidoglycan Recognition Protein (PGRP)-LB, apparently plays an important role in the density regulation of *Wigglesworthia* in the tsetse host.

Keywords : Wigglesworthia, tsetse flies, mutualism, vector control

#### Comparative genomics of insect endosymbionts

<u>Klasson L.</u>¹, Tamarit D., Ellegaard K.M.¹, Näslund K.¹, Bourtzis K.^{2,3} and Andersson S.G.E.¹

¹Molecular Evolution, Department of Cell and Molecular Biology, Uppsala University, Sweden; ²Department of Environmental and Natural Resources Management, University of Western Greece, Greece; ³Biomedical Sciences Research Center Al. Fleming, 16672 Vari, Greece - lisa.klasson@icm.uu.se

Many endosymbionts cannot be cultivated in artificial media and their DNA can only be obtained after isolation of large numbers of bacterial cells directly from the hosts. Not surprisingly, it may take years to get enough DNA for sequencing. We have developed a method to isolate, purify and amplify endosymbiont DNA from small quantities of starting material. We applied the method to the isolation, amplification and sequencing of Wolbachia strains infecting Drosophila simulans. Endosymbionts were isolated from the eggs of flies reared in our laboratory and after DNA amplification we sequenced the genomes of Wolbachia strains wHa and wNo, which belong to supergroup A and B, respectively. The comparative analysis of these two genomes with those already sequenced from supergroups A and B has revealed a massive exchange of genes among strains within supergroups as well as a few examples of genetic exchange across the two supergroups. Additionally, we have applied these methods to the study of endosymbionts isolated from aphids and other insects collected from the environment. The endosymbionts contained inside these insects were dissected under microscopy, and their DNA was amplified and sequenced. We have identified both primary and secondary endosymbionts. We have selected samples from aphids containing the secondary endosymbiont Hamiltonella defensae for a more detailed comparative genomics study, with a focus on putative virulence genes. We discuss the role of recombination and gene exchange for the evolution of insect endosymbionts.

# *'Candidatus Cardinium* hertigii' - a reproductive manipulator with a role in host nutrition

Penz T.¹, Schmitz-Esser S.², Kelly S.E.³, Müller A.², Woyke T.⁴, Malfatti S.A., Hunter M.S.³ and Horn M.¹

¹Department of Microbial Ecology, University of Vienna, Althanstrasse 14, 1090 Vienna, Austria; ²Institute for Milk Hygiene, University of Veterinary Medicine Vienna Veterinärplatz 1, 1210 Vienna, Austria; ³Department of Entomology, University of Arizona, 85721 Tucson, AZ, USA; ⁴DOE Joint Genome Institute, 94598 Walnut Creek, CA, USA horn@microbial-ecology.net

Bacterial symbionts of insects have evolved sophisticated strategies to manipulate their host's reproduction to ensure survival and spread within the host population. This may lead to sex role reversal and might even contribute to host speciation. Cardinium hertigii, a member of the Bacteroidetes and maternally transmitted symbiont of the parasitic wasp *Encarsia pergandiella* is the only known reproductive manipulator outside of the Alphaproteobacteria, which causes cytoplasmic incompatibility (CI) constraining crosses between uninfected females and infected males. Here we report the complete genome sequence of Cardinium hertigii strain Texas. Comparative analysis with the next closest sequenced relative, the amoeba symbiont Amoebophilus asiaticus, provides insights into the evolution of symbiosis in the Bacteroidetes. The metabolically restricted genome of Cardinium lacks all major biosynthetic pathways but harbors a complete biotin biosynthesis pathway, which suggests a role of Cardinium in host nutrition. Cardinium lacks known protein secretion systems but encodes a putative new, phage derived secretion system distantly related to the antifeeding prophage of the entomopathogen Serratia entomophila. Comparison of the Cardinium genome with the genomes of other CI inducing reproductive manipulators, the Wolbachia pipientis strains wMel, wRi and wPip, provides a unique opportunity to pinpoint conserved proteins mediating host cell interaction. This allowed us to propose a set of candidate proteins possibly involved in CI. Taken together, the reproductive manipulator Cardinium shows genome characteristics of known obligate, mutualistic symbionts of insects as well as of parasitic CI inducing Wolbachia strains, but seems to be on the path to become a long-term obligate symbiont.

Keywords : cytoplasmic incompatibility, evolution, phage

#### Arsenophonus genomes

Novakova E.^{1,2}, Hypsa V.¹, Moran N.A.², Lehane³ M., Hurst G.D.D.³ and <u>Darby A.C.³</u>

¹Faculty of Science, University of South Bohemia, and Institute of Parasitology, Biology Centre, ASCR, v.v.i., Branisovka 31, 37005 Ceske Budejovice, Czech Republic; ²Department of Ecology and Evolutionary Biology, Yale University, 300 Heffernan Dr, West Haven, CT 06516-4150, USA; ³ Institute of Integrative Biology, University of Liverpool, Crown Street, Liverpool, L69 7ZB, United Kingdom - acdarby@liv.ac.uk

Arthropods commonly have symbiotic relationship with maternally inherited bacteria. These relationships are diverse, and include obligate mutualism, facultative benefit to the host, and reproductive parasitism. Our understanding of patterns of genomic evolution under different forms of symbiosis has generally relied on contrast to either free living relatives or distantly related symbionts. The genus *Arsenophonus* is unusual in containing symbionts whose interactions with their hosts are diverse in themselves, and therefore allows comparison of genome form between closely related symbionts that share a recent common ancestor but display very different interactions with their host.

We demonstrate that Arsenophonus lineages adapted to different lifestyles also show substantial differences in important genomic traits. These differences reveal general tendencies along the hypothetical evolutionary continuum from almost "standard" bacterial genomes in parasites and facultative symbionts to highly modified and degenerated genomes in obligate mutualists. The most typical manifestations of these tendencies are massive gene loss, compositional bias and economization of the genome with decreasing frequency of horizontal gene transfer.

Keywords: Arsenophonus, Genome Evolution, transmission

/	)
1	
$\vdash$ $-$	 
1	
1	
1	
1	
<u> </u>	 
<u> </u>	 
$\vdash$ $-$	 
1	
1	
1	
$\vdash$ $-$	 
1	
1	
1	
1	
1	
1	
1	
1	
1	 i a
1	
1	
1	
1	
1	
1	
\	/
\	/
\	
	/

### The story of communities affecting communities

#### Vavre F.

Laboratory of Biometry and Evolutionary Biology, UMR CNRS 5558, University Lyon 1, Villeurbanne, FRANCE. fabrice.vavre@univ-lyon1.fr

Arthropod symbiosis has long been studied by considering only a "one hostone symbiont" approach. However, bacterial symbionts directly or indirectly affect the interactions of the host with other species within a community, and multiple symbionts often share the same hosts. Integrating these multiple interactions at different levels of integration is a challenging task, but is clearly a way to pursue to better understand the functioning and evolutionary trajectory of host-symbiont interactions and to use symbionts for applied perspectives.

In this talk, I will briefly review some concepts showing that the community level may play a fundamental role in shaping symbiosis at different levels, from populations to genomes. The whitefly *Bemisia tabaci* will be used to illustrate this and to show progresses that have been made on this system by gathering different teams from the COST action.

One conclusion is that insects' symbiotic complements are dynamic communities that affect and are affected by the communities in which they are embedded.

Keywords: Multiple infections, competition, cooperation, genomics, Bemisia tabaci

/	)
<u> </u>	 
1	
$\vdash$ $-$	 
1	
1	
1	
1	
<u> </u>	 
<u> </u>	 
$\vdash$ $-$	 
1	
1	
1	
$\vdash$ $-$	 
1	
1	
1	
_	 
1	
1	
1	
1	
1	
1	 i a
1	
1	
1	
1	
1	
1	
\	/
\	/
\	
	/

# Nitrogen recycling in the aphid/Buchnera symbiosis: the role of the host cell

<u>Macdonald S.J.</u>¹, Lin G.G.², Russell C.W.², Douglas A.E.² and Thomas G.H.¹

¹Department of Biology (Area 10), University of York, York, UK, & ²Department of Entomology, Comstock Hall, Cornell University, Ithaca, NY, USA. sandyjmacdonald@gmail.com

The pea aphid/Buchnera system is a well characterized symbiosis, where the process of nitrogen recycling operates to enable the aphid to reuse a limited supply of nitrogen via its bacterial partner for synthesis of essential amino acid (EAA). Using in silico metabolic modelling with complementary transcriptomic, proteomic and metabolite profiling we have investigated nitrogen recycling in this model symbiotic partnership. The data suggest that, contrary to expectations, aphid waste ammonia is recycled predominantly by the host cell (bacteriocyte) and not Buchnera. This recycling of host waste ammonia is mediated through the terminal transaminase reactions of four EAA biosynthesis pathways which occur within the bacteriocytes and not the Buchnera. Also, our data suggest that this ammonia is generated directly within the bacteriocytes as a consequence of the adaptation of bacteriocyte metabolism to provide metabolic precursors to the Buchnera using non-essential amino acids as substrates. The modeling also reinforces additional metabolic interactions between the two partners over and above EAA biosynthesis such as complementary purine metabolism and pantothenate biosynthesis.

Keywords: Aphid, Buchnera aphidicola, flux balance analysis, bacteriocytes, essential amino acids

## *Sodalis glossinidius*, a symbiotic Trojan horse against a tsetse fly transmitted parasite - the trypanosome

Van Den Abbeele J.

Institute of Tropical Medicine Antwerp, Dept. Biomedical Sciences, Nationalestraat 155, B-2000 Antwerp, Belgium, jvdabbeele@itg.be

African trypanosomiasis is a devastating vector-borne parasitic diseases in human and animals caused by protozoan parasites of the genus Trypanosoma. These parasites are biologically transmitted by an obligatory blood feeding insect, the tsetse fly (Glossina sp.) in which the parasite undergoes a complex life cycle with several rounds of differentiation and proliferation, orchestrated by a largely unknown molecular parasite-vector crosstalk. Sodalis glossinidius is a maternally inherited gram-negative bacterial endosymbiont of the tsetse fly that can be found both inter- and intracellularly in the tsetse fly midgut, muscle, fat body, milk glands, and salivary glands. Given the close proximity of S. glossinidius to the different insect tissues where the trypanosome parasites reside and the fact that this symbiotic bacteria can be cultured and genetically modified in vitro, this bacterium is considered as a potential in vivo drug delivery vehicle to control T.congolense and T.brucei development in the fly. In this context, we recently demonstrated the expression and extracellular release of functional trypanosome-targeting Nanobodies by genetically modified Sodalis symbionts. Furthermore, these Sodalis strains were not affected in their growth, suggesting that they may be competitive with endogenous microbiota in the midgut environment of the tsetse fly. These data strengthen the view that a paratransgenic approach using Sodalis as a Trojan horse for trypanosome-targeting Nanobody delivery has a realistic potential to generate parasite-resistant tsetse flies. Moreover, this approach could also open a new avenue to unravel the molecular determinants of the specific parasite-vector crosstalk during development in the tsetse fly. This innovative Nanobody delivery through bacterial symbionts could also be applicable for paratransgenesis in other vector-pathogen systems.

Keywords: tsetse fly, trypanosome, Sodalis glossinidius, paratransgenesis, nanobodies

### Immunity and symbiosis in weevils

### Heddi A.

INSA-Lyon, UMR203 BF2I, INRA, Biologie Fonctionnelle Insectes et Interactions, Bâtiment Louis-Pasteur 20 Avenue Albert Einstein, F-69621 Villeurbanne, France Abdelaziz.Heddi@insa-lvon.fr

Mutualistic associations between animals and bacteria are widespread in nature and are particularly common in insects that exploit unusually restricted nutritional resources. Among symbiotic insects, many are serious pests of crops or vector of animal diseases. Insect intracellular bacteria (endosymbionts) are transmitted vertically for millions of years and provide nutrient supplementation to their hosts, thereby improving their physiology, their adaptive capabilities and their invasive power.

However, maintaining the beneficial nature of this long-term relationship constrains both the host and the symbiont to adaptive changes in their interaction. Evolutionary data have shown that upon establishment, insect endosymbionts experience drastic gene deletions and mutations, which affect among other genes involved in bacterial virulence and host tolerance. Contrariwise, knowledge on how host immune system is regulated to tolerate cooperative bacteria remains scarce and mainly limited to intestinal lumen microbiota and few extracellular bacterial associations.

Insect obligate associations have selected an "endosymbiont compartmentalization" strategy to control endosymbiont space and number. In addition to germ cells, from which endosymbionts are maternally transmitted to offspring, bacteriocytes are the only cells that tolerate endosymbionts and permit their growth and division. Bacteria escaping bacteriocytes face in the hemolymph the activation of a systemic immune response that produce effectors similar to those expressed against pathogens.

Specific molecular features of the weevil bacteriocytes were approached with transcriptomic analyses. Bacteriocyte cells were shown to undergo a modulated immune response. Except the coleoptericinA (ColA) antimicrobial peptide (AMP), AMP encoding genes are slightly (thought not) expressed within the bacteriocytes, probably to allow symbiont maintenance. Functional investigations showed that ColA selectively targets endosymbionts within the bacteriocytes and regulates their growth through the inhibition of cell division. Silencing the colA gene with RNAi resulted in a decrease in size of the giant filamentous endosymbionts, which then escaped from the bacteriocytes and spread to insect tissues.

While AMPs have commonly been linked with microbe clearance, these findings demonstrate that endosymbiosis benefits from ColA and suggest that long-term host-symbiont coevolution might have shaped immune effectors for symbiont maintenance.

Keywords: Symbiosis, immune system, endosymbiont control, evolution

																												$\backslash$
/																												
																												1
	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	_	-
	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	
L _	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_
<u> </u>	_	—	—	_	_	—	_	—	—	—	—	—	—	—	—	—	_	—	_	—	—	_	—	—	—	_	—	_
$\vdash$ $-$	_	_	—		—	_	—	—	—	—	_	_	—	—	—	—	—	—	—	—	—	_	—	—	_	—	—	$\neg$
1																												
1																												
1																												
1																												
1																												
	-	_	_	_	_	_	_	_	—	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	
		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	
	_	_	_		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_
<u> </u>	_	_	_			_	_	_	_	—	_	_	_	—	_	_	_	_	—	_	_	_	_	_	_	_	_	_
	_	—	—	_	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	-
	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_
1																												
1																												
1																												
1																												
		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	
	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_		_	_	_	_	_	
1																												
1																												
L _	_		_	_	_	_		_	_	_	_		_	_	_			_		_	_	_	_	_	_	_	_	
								-	-			_		_		-				_	_	_	_	-		_	_	
1																												
1																												
1																												
1																												
						_		_	_	_	_	_	_	_	_	_				_	_		_		_	_	_	
1																												
1																												
1																												
1																												
1																												
			_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	-		_
																											Â	·
																										1		
																										X		
																									11			
																									$\mathbb{Z}$			
1																								1		/		
1																						~	~	$\sim$				
																						-		-				
\																												
																												/
$\sim$																											/	

## From insect pathogen to obligate symbionts: what's in between?

#### Latorre A.

Instituto Cavanilles de Biodiversidad y Biología Evolutiva. Grupo de Genética Evolutiva. C/ Catedrático José Beltrán nº 2. 46980 Paterna, Valencia, Spain. amparo.latorre@uv.es

A critical issue in evolutionary biology is to find traits or organisms that provide evidence of the transition from one lifestyle to another, no matter how gradual the process may be. The evolutionary history of intracellular symbiosis involving the transition of bacteria from free-living to obligate lifestyle reveals the existence of different types of bacteria, ranging from mere facultative to obligatory symbionts. Bacterial species with different strains having both types of traits, as it is the case of *Serratia symbiotica* in aphids, represent an important "missing link" in the history of this process.

Aphids are insects that feed on plant-sap, an unbalanced nitrogen/carbon diet deficient in a number of nutrients, such as essential amino acids, which aphids cannot synthesize and are provided by Buchnera aphidicola, their primary endosymbiont. In addition to B. aphidicola, some aphid populations harbor additional facultative (or secondary) symbionts that are not required for growth or reproduction, being S. symbiotica the most occurring in aphids. Nowadays, the genomes of B. aphidicola from five different aphid species have been fully sequenced, being the one from C. cedri, with only 416 kb, the smallest known Buchnera genome¹. Its sequencing and functional analysis revealed that this bacterium had lost part of its symbiotic role as it was not able to synthesize tryptophan. More recent studies showed that the biosynthesis of tryptophan is shared with the symbiont S. symbiotica SCc, which coexists with B. aphidicola in this aphid, and thus cannot be considered as facultative. The whole genome sequencing of S. symbiotica SCc² reveals an endosymbiont in a stage that is closer to an obligate endosymbiont, such as B. aphidicola from the aphid Acyrthosiphon pisum (with a genome size of 640 kb), than to the facultative S. symbiotica from the same aphid. Moreover, the comparative genomic study of five strains of Serratia, three free-living and two endosymbiotic ones, has let us to disentangle the evolutionary steps undergone from an insect pathogen to an obligate symbiont.

¹Pérez-Brocal, V. et al. 2006, Science, 314:312-313. ²Lamelas, A. et al. 2011, PLoS Genetics 7(11) e1002357

# Immune response of the ant *Camponotus floridanus* against pathogens and its obligate mutualistic endosymbiont

Ratzka C.¹, Feldhaar H.² and Gross R.¹

¹University of Wuerzburg, Department for Microbiology, Germany; ²University of Bayreuth, Department for Animal Ecology, Germany carolin.ratzka@biozentrum.uni-wuerzburg.de

Bacterial endosymbionts are very common in insects and can confer several fitness benefits to their hosts. Insect hosts face the problem of having to maintain such mutualistic bacteria while staging an immune response towards pathogens upon infection. Camponotus floridanus ants harbour the obligate endosymbiont Blochmannia floridanus in specialized midgut cells and ovaries. The distribution of bacteriocytes and of Blochmannia endosymbionts varies strongly during the holometabolous life cycle of the ant hosts [1]. Especially, during metamorphosis the proportion of endosymbiont-bearing midgut cells increases strongly and peaks in late pupal stages, where the entire midgut is transformed into a symbiotic organ. After eclosion of workers the symbiosis degenerates as bacterial numbers decrease with age of workers. It was hypothesized that hosts could regulate the number of endosymbionts present in their tissues via the innate immune system [2]. A quantitative gene expression analysis revealed distinct expression patterns of some immune genes according to developmental stage and tissue. Particularly, two pattern recognition receptor genes, PGRP-LB and PGRP-SC2, reached maximal expression levels in the pupal midgut tissue at the time, when the highest number of Blochmannia is present in this tissue.

A quantitative analysis of immune gene expression revealed different expression kinetics of individual factors and also characteristic expression profiles after injection of gram-negative and gram-positive bacteria. Likewise, *B. floridanus* injected into the hemocoel elicited a comparable immune response of its host *C. floridanus*. Thus, the host's immune system may prevent colonization of tissues other than the bacteriocytes by *Blochmannia* and may play a role in the regulation of the number of bacteria present within bacteriocytes.

1. Stoll S, Feldhaar H, Fraunholz MJ, Gross R (2010) Bacteriocyte dynamics during development of a holometabolous insect, the carpenter ant Camponotus floridanus. BMC Microbiol 10: 308.

2. Ratzka C, Liang C, Dandekar T, Gross R, Feldhaar H (2011) Immune response of the ant Camponotus floridanus against pathogens and its obligate mutualistic endosymbiont. Insect Biochem Mol Biol.

	-																											~	
/																													
		_	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	-
		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	
		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	—	_	_	_	_	_	_	_	_	_
		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	—	_	_	_	_	—	_	_	—	_
	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	
		-	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	
		_	_	_	_	_	_	_	_	_	_	_	_		_	_	_	_	_	_	_	_	_	_		_	_	_	
		_	_		_	—	_	_	_		_	_	_	_		_	_	_		—	_	_	_			_	_	_	_
		_	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	-
		_	_	_	_	_	_	_	—	_	_	_	_	_	_	_	_	_	_	—	_	_	_	_	—	_	_	_	_
	_									_					_														
		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_
		_	_	—	—	—	—	_	—	—	—	—	—	—	—	—	_	—	—	—	—	_	_	—	—	—	_	—	-
		-	—	—	—	—	—	_	—	—	—	_	_	_	—	—	—	—	—	—	—	_	_	—	—	—	_	—	$\neg$
										_						_	_	_				_	_	_	_	_			
		_		_	_									_			_	_		_	_	_	_			_	_		1
					_	-	_	_	-				-	_	_	_	_	_	_	_	_	_	_	_	-	_	-	_	
1			_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_					_	_			1	1
																											10		
																										.11	7		
																										1	1		
																							-	~	r	-			
1																							~	4 4					
\																													/
\																													/
	_																											/	1
				_	_	_	_	_											_	_	_	_							

## Symbionts mediate coevolution between aphid hosts and parasitoids

Vorburger C.

Institute of Integrative Biology, ETH Zürich, 8092 Zürich, Switzerland & EAWAG, Swiss Federal Institute of Aquatic Science and Technology, 8600 Dübendorf, Switzerland - christoph.vorburger@eawag.ch

The present understanding of host-parasite coevolution is based on models that assume a direct genetic interaction between host and parasite. This assumption has been challenged by recent empirical findings, showing that hosts often rely on 'helpers' in the form of microbial endosymbionts for defence against parasites. The best-studied case are aphids, in which much of the variation in resistance to parasitoids is due to facultative endosymbiotic bacteria. Using the example of the black bean aphid, Aphis fabae, and its parasitoid Lysiphlebus fabarum, I illustrate how defensive endosymbionts alter the reciprocal selection between host and parasitoid and thus mediate coevolution. They increase the variation available to selection and make the host-parasite interaction more specific as a consequence of strong genotype-bygenotype interactions between the parasitoids and the aphids' endosymbionts. Protection by symbionts does not come for free to the host. Infected aphids suffer from a reduced lifespan and lifetime reproduction in the absence of parasitoids, and the magnitude of this costs is again determined by a genotypeby-genotype interaction, here between aphids and their symbionts. I refer to this complex three-way interaction as 'symbiont-mediated coevolution', and I highlight the need for new models to understand this intriguing process.

Keywords: Coevolution, genotype x genotype interaction, Hamiltonella defensa, resistance, symbiosis

_			 																_	
																				$\backslash$
(																				
		 	 	 -	—	—	—	—	—				_	—	—	—	—	—	_	-
	·	 	 	 —	—	—	—	—	—				—	—	—	—	—	—	—	-
	·	 	 	 _	_	_	_	_	_				_	_	_	_	_	_	_	_
L		 	 	 _	_	_	_	_	_				_	_	_	_	_	_	_	_
L_		 	 _	 	_	_	_	_	_	_				_	_	_	_	_	_	
		 _	 			_	_	_	_		_	_		_	_	_	_		_	1
		 	 	 _	_	_	—	_	_				_	_	—	—	_	_	_	-
	·	 	 	 —	—	_	—	—	_				_	—	—	—	—	—	—	-
	·	 	 	 _	_	_	_	_	_				_	_	_	_	_	_	_	_
		 	 	 _	_			_	_				_	_	_		_	_	_	_
		 	 	 _	_	_	_	_	_				_	_		_	_	_		
F	·	 	 	 _	_	_	_	_	_				_	—	_	—	—	_	_	
	·	 	 	 _	—	_	—	—	_				_	—	—	—	—	_	_	-
⊢	·	 	 	 _	_	_	—	_	_	_ ·			_	_	_	_	_	_	_	_
		 	 _	 	_		_	_	_	_			_			_	_		_	
	_	 _	_	 _	_	_	_	_	_	_				_	_	_	_	_	_	
		 	 _	 	_	_	_	_	_	_			_	_	_	_	_	_	Å	~
																		/		
																	11	1		
																		1		
															$\sim$	~				
			 	 																/

## Disease epidemiology in arthropods is altered by the presence of non-protective symbionts

Ryder J.J.^{1,2}, Hoare M.J.¹, Pastok D.¹, Bottery M.¹, Boots M.², Atkinson D.¹, Knell R.J.³ and <u>Hurst G.D.D.¹</u>

¹Institute of Integrative Biology, University of Liverpool, Crown Street, Liverpool L69 7ZB, U.K.; ²Centre for Ecology & Conservation, Biosciences, College of Life & Environmental Sciences University of Exeter, TR10 9EZ, U.K.; ³School of Biological and Chemical Sciences, Queen Mary, University of London, Mile End Road, London E1 4NS, U.K. - G.Hurst@liverpool.ac.uk

Inherited microbial symbionts can profoundly alter the susceptibility of their arthropod hosts to pathogen and parasite infection. A wider range of symbionts influence host demography, potentially modifying host disease risk through altering transmission opportunities. We tested whether the demographic effects produced by symbionts were sufficient to alter disease dynamics, examining the impact of a sex ratio distorter on the dynamics of a sexually transmitted infection (STI). We examined this in the two spot ladybird Adalia bipunctata, which carries a mite STI and a range of male-killing bacteria, and compared STI dynamics between populations where the male-killer was common and ones where it was rare. We observed that presence of Spiroplasma male-killers was associated with male-biased epidemic spread of the STI, driven by the strongly female-biased sex ratios produced by Spiroplasma male-killers. There were no individual effects of Spiroplasma infection on susceptibility to mite infection, and conclude therefore that the differences in STI epidemiological differences arose solely as a consequence of the male-killer on population sex ratio. We conclude that understanding and predicting parasite dynamics in arthropods requires incorporation of the effects of symbionts and that the range of symbionts that are likely to alter parasite epidemiology is much wider than previously envisaged, because it will include those that impact host demography without altering susceptibility.

																												$\backslash$
/																												
																												1
	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	-
	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	
L _	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_
<u> </u>	_	—	—	_	_	—	_	—	—	—	—	—	—	—	—	—	_	—	_	—	—	_	—	—	—	_	—	_
$\vdash$ $-$	_	_	—		—	_	—	—	—	—	_	_	—	—	—	—	—	—	—	—	—	_	—	—	_	—	—	$\neg$
1																												
1																												
1																												
1																												
1																												
	_	_	_	_	_	_	_	_	—	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	
		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	
	_	_	_		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_
<u> </u>	_	_	_			_	_	_	_	—	_	_	_	—	_	_	_	_	—	_	_	_	_	_	_	_	_	-
	_	—	—	_	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	-
	_	_	_		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_
1																												
1																												
1																												
1																												
		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	
	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_		_	_	_	_	_	
1																												
1																												
L _	_		_	_	_	_		_	_	_	_		_	_	_			_		_	_	_	_	_	_	_	_	
								-	-			_		_		-				_	_	_	_	-		_	_	
1																												
1																												
1																												
1																												
						_		_	_	_	_	_	_	_	_	_				_	_		_		_	_	_	
1																												
1																												
1																												
1																												
1																												
			_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	-		_
																											Â	·
																										1		
																										X		
																									11			
																									$\mathbb{Z}$			
1																								1		/		
1																						~	~	$\sim$				
																						-		-				
\																												
																												/
$\sim$																											/	

## Experimental evolution of virulence in a symbiosis: from vertical to horizontal transmission

Le Clec'h W., Bertaux J., Bouchon D. and Sicard M.

Laboratoire Ecologie et Biologie des interactions UMR 7267 CNRS, Université de Poitiers, France. mathieu.sicard@univ-poitiers.fr

Wolbachia, may be the most widespread endosymbiotic bacteria in arthropods, are vertically transmitted from mother to offspring. In their native hosts, they are weakly virulent except for one virulent strain in Drosophila called wMelpop. This global feature in term of virulence is coherent with theoretical works predicting evolution towards limited virulence for vertically transmitted symbionts. In this talk, I will present data that indicate that Wolbachia can evolve quickly towards strong virulence when horizontally transmitted. By using Armadillidum vulgare's hemolymph, that naturally contains immune cells infected by Wolbachia (wVulC), as a vector of infection from one individual to another, we forced Wolbachia to evolve under the major constraint of horizontal transmission. After only few horizontal passages via hemolymph injection, the number of *Wolbachia* recorded in the receiving animals increased dramatically compared to the beginning of the experiment. Then, the number of bacteria infecting receiving hosts still increased at each passage. In parallel, the mortality of the naïve animals that received the bacteria also strongly increased starting from 0% at the first passage to reach 90% after only 6 selection passages. Our results indicate that Wolbachia can rapidly adapt their virulence due to a new transmission mode and become deadly pathogens.

Keywords: Virulence, Evolution, Transmission mode, Symbiont, Pathogens

## Using *Wolbachia* infections to control dengue transmission

### O'Neill S.L.

### School of Biological Sciences, Monash University, VIC 3800 - scott. oneill@monash.edu

Dengue fever is an emerging insect-transmitted disease causing illness in more than 50 million people each year. Currently there are no effective treatments beyond prevention measures targeting the main mosquito vector. Our group is examining the potential use of *Wolbachia* as a novel method to interfere with dengue transmission. This work is now moving from basic bench studies into open field trials in Australia, Vietnam and Indonesia. The presentation will give an overview of current understanding of *Wolbachia*mosquitio-pathogen interactions as well as the current status of the applied program.

/		
		1
L		
L		
L .		
$\vdash$ $ -$		
		A
		r
	~~~~	
1		1
\		/
\		
		/
		_

Is the "multicoloured" microbiome associated with insect vectors of plant pathogens a potential resource for disease control?

Daffonchio D.

DISTAM, University of Milan, via Celoria 2, 20133 Milan Italy, daniele. daffonchio@unimi.it

Different types of symbiotic microorganisms have been reported in sapfeeding insects like leafhoppers and planthoppers that are important for their capability of transmitting plant pathogens to plants. Such symbionts have different prevalence and topology in the host body, undergo complex transmission pathways both vertical and horizontal and support many important functions for the hosts. By considering as examples the planthopper Hyalesthes obsoletus (Hemiptera: Cixiidae) and the leafhopper Scaphoideus titanus (Hemiptera: Cicadellidae) both vectors of phytoplamas pathogenic to grapevine, we show different features of these symbionts and discuss their potential for blocking disease transmission. In H. obsoletus the high infection rates of the Bacteroidetes "Candidatus Sulcia muelleri" the Gammaproteobacterium "Candidatus Purcelliella pentastirinorum" and the Betaproteobacterium "Candidatus Vidania fulgoroideae" suggest a tripartite primary symbiosis with the host. These symbionts are not alone being associated to the intracellular bacteria "Candidatus Wolbachia pipientis", "Candidatus Rickettsia sp.", and "Candidatus Cardinium hertigii" that are localized in different organs and tissues as revealed by fluorescent in situ hybridization. The diversity of the symbiotic processes is highlighted in S. titanus where Cardinium is capable of colonizing very different tissues besides the female gonads. From the salivary glands Cardinium can be released in the feeding medium and the plant tissues, to be up taken by other individuals in an horizontal transmission mode associated to a co-feeding habit. The diversity of the biological processes mediated by such a diverse range of symbionts is a potential source of novel approaches for blocking pathogenic microorganisms during their transmission to the plant.

Keywords: mutualistic symbiosis, Scaphoideus titanus, Hyalesthes obsoletus, symbiont transmission

Mosquitoes-symbionts interactions: from complex relationships to biotechnological applications

<u>Guido F.</u>

School of Biosciences & Biotechnology, University of Camerino, Camerino (MC) Italy - guido.favia@unicam.it

Several tens of mosquitoes out of around 3500 species described to date are vectors of pathogens of great interest in public health. Mosquitoes show a great ability to adapt to very different habitats where they play relevant ecological roles. In fact, they are present all around the world. It is very likely that the mosquito microbiota have provided the mosquito a great capacity to adapt to many different environments. Thus, a better understanding of the mosquito-microbiota relationships may have a great impact in a better knowledge of some traits of mosquito biology and in the development of innovative mosquito-borne disease-control strategies aimed to reduce mosquito vectorial capacity and/or inhibiting pathogen transmission. In this context, since some years the unit of Molecular Entomology of the University of Camerino has started a study for the identification of mosquito symbionts. In particular we have focused our interest on two of them: 1) The acetic acid bacterium Asaia and 2) The yeast Wickeramomyces anomalus. Asaia is a Gram-negative, found in some mosquito species including several malaria vectors and dengue virus vectors. Asaia is the most prevalent bacteria in both natural and lab mosquito populations and its prevalence often reaches 100%, detected in high numbers in the midgut, salivary glands, and reproductive organs. For all these reasons Asaia could has been proposed to express anti-pathogen(s) molecules directly in the midgut of the mosquito to exert an inhibitory effect against pathogens. Furthermore, its presence in the reproductive organs permits a vertical transmission route that occurs both maternally and paternally thus providing the basis for the introduction of engineered bacteria into mosquito populations in the field. Experiments performed with engineered bacteria expressing fluorescent proteins has provided a proof of principle to the feasibility of using Asaia to express anti-parasite effectors to control malaria and others mosquito-borne diseases At the present we have focusing on the "function(s) that Asaia is exerting on the mosquito biology. Asaia in mosquito larval development has been previously shown to reduce the time required from the larvae to develop in pupae. Currently we are assessing the impact of Asaia on adult mosquito vitality. W. anomalus was detected at all the developmental stages of both malaria and dengue mosquito vectors where it localizes in the midgut and reproductive organs. The gut and gonads may provide nutrients for yeasts, thus representing optimal niches for symbionts. The finding that W. anomalus associates with some mosquito vectors of several human parasites led to the proposition to use W. anomalus to control mosquito-borne diseases. Moreover, some strains of W. anomalus produce killer toxins with antimicrobial activities against several human pathogens including arthropod-transmitted parasites such as Leishmania spp.. Preliminary observations indicate that the yeast strains isolated from a mosquito also produce killer toxins thus suggesting the possible use of W. anomalus in the control of mosquito-borne diseases. Studies for the biochemical characterisation of this protein are in due course. In conclusion, the field of mosquito symbiosis is providing an important contribution to the knowledge of the biology of mosquitoes. Curiositydriven studies have lead to new insight into the biology of facultative symbionts likely affecting relevant traits of mosquito vectors. Genetic transformation of the symbionts with genes encoding for anti-pathogen effector molecules as well as the use of killer toxins produced by mosquito are interesting field of application, that could contribute to the set-up of integrated control strategies.

/		
		1
L		
L		
L .		
\vdash $ -$		
		A
		r
	~~~~	
1		1
\		/
\		
		/
		_

### Symbiotic control of honeybee pathogens

Hamdi C.^{1,2}, Balloi A.³, Essanaa J.¹, Abdi K.², Crotti E.³, Najjari A.¹, Belghuith K.², Barbouch N.⁴, Alma A.⁵, Daffonchio D.³, Boudabous A.¹ and <u>Cherif A.^{1,2}</u>

¹Laboratoire Microorganismes et Biomolécules Actives, Faculté des Sciences de Tunis, Université de Tunis El Manar, 2092, Tunis, Tunisia; ²Laboratooire Biotechnologie et Valorisation des Bio-Géo Ressources, Institut Supérieur de Biotechnologie de Sidi Thabet, Université de la Manouba, 2020, Ariana, Tunisia; ³Dipartimento di Scienze e Tecnologie Alimentari e Microbiologiche, Università degli Studi, 20133, Milano, Italy; ⁴Laboratoire de Zoologie et Apicultures, Institut National Agronomique de Tunis, 43 Avenue Charles Nicolle, 1082 Tunis Mahrajène, Tunisia; and ⁵Di.Va.P.R.A. Entomologia e Zoologia applicate all'Ambiente, Università degli studi di Torino, Italy - cherif.ameur@ gmail.com

Honeybees are hosts for a large diversity of microorganisms, including commensal, probiotic and pathogenic bacteria. *Paenibacillus larvae* is the primary *bacterial pathogen* that cause a *serious disease* called American foulbrood (AFB). For controlling AFB and other bacterial brood diseases, antibiotics were extensively used which has led the appearance of pathogen resistance and the risk of contamination of hive products. For these reasons, the pressing public concern pointed out the need of finding alternative control strategies; one of these could be based on the symbiotic control approach.

In our laboratory, we are interested in studying the composition of bacterial communities in honeybee with cultivation-dependent (isolation experiments) and independent techniques (DGGE and ARISA). Ribosomal genes-based methods including ARISA and PCR-DGGE on total DNA from the honeybee gut showed remarkable differences between AFB-diseased and healthy larvae. In AFB-diseased larvae a community less complex than in the healthy ones occurred, in which *P. larvae* appeared to thrive and dominate. In healthy individuals, bacteria mainly represented by *Lactobacillus* and the acetic acid bacteria (AAB) *Saccharibacter*, *Acetobacter* and *Gluconobacter* have been detected. Isolation experiments conducted on AFB-diseased honeybees allowed the establishment of a large collection of *P. larvae* (210 isolates) that showed a previously unrecognized high intraspecific diversity. The isolation of bacterial symbionts from healthy larvae left a large collection of *Bacillaceae*, *Lactobacilli* and AAB of the genus *Saccharibacter*.

Several of these honeybee-associated bacteria showed high capacity to inhibit the growth of P. larvae strains in-vitro. These antagonists belonged to the Bacillus, *Brevibacillus* and *Lactobacillus* genera. The efficiency of the active bacteria was tested on in-vitro reared larvae and then at colony level (in the field). Our overall results showed the ability of the active strains in preventing from P. larvae infestation in larval tissues (in vivo) and to counteract the development of AFB disease at colony level after several applications on bee hives. For other bee diseases, further symbiotic control experiments are currently in progress.

### Tephritid symbionts and their applied potential

Yuval B., Gavriel S. and Jurkevich E.

#### Department of Entomology, Hebrew University of Jerusalem. Rehovot, Israel - yuval@agri.huji.ac.il

Tephritid fruit flies (Diptera: Tephritidae), harbor a diverse community of bacteria in their digestive system. Members of the Enterobacteriaceae form the dominant populations in the medfly (Ceratitis capitata) gut, and contribute to various physiological and behavioral parameters related to fitness. Microbial communities in mass rearing strains used for the sterile insect technique (SIT) differ from wild flies, and irradiation further impoverishes the microbial community. We tested the hypothesis that inoculation of sterile flies with members of the original bacterial community results in enhanced competitiveness. Specifically we determined how feeding on diets enriched with *Klebsiella oxytoca* affect the ability of sterile males to compete for wild females against wild males, their ability to inhibit female receptivity, and their survival. We found that enriching the sterile male diet with K. oxytoca significantly improved mating competitiveness in the laboratory and in field cages. In addition, bacterially enriched sterile males inhibited female receptivity more efficiently than sugar fed males and survived longer duration of starvation. We conclude that inoculating mass reared sterile flies with bacteria prior to their release is a valid approach to improve the efficacy of the SIT.

Keywords: Symbiosis, Tephritidae, Sterile Insect Technique

/	)
1	
$\vdash$ $-$	 
1	
1	
1	
1	
<u> </u>	 
<u> </u>	 
$\vdash$ $-$	 
1	
1	
1	
$\vdash$ $-$	 
1	
1	
1	
1	
1	
1	
1	
1	
1	 e de la
1	
1	
1	
1	
1	
1	
\	/
\	/
\	
	/

## The potential of symbionts in improving biological control agents

### T. Groot and K. Bolckmans

#### Research & development, Entomology department, Koppert B.V. Veilingweg 17, P.O. Box 155, 2650 AD Berkel en Rodenrijs, Nederland

With the increased interest in the interactions between arthropods and their symbionts has come the realization that symbionts can be used to control arthropod pests. Several symbiont based control strategies are currently being developed, but in most cases still quite some time will pass before these methods will be implemented on a large scale. However, even if symbionts aren't the basis of the control strategy, they still can be important in biological control strategies. For example, symbionts that induce parthenogenesis will allow more efficient production of a parasitoids and better control in the field. In contrast, cytoplasmic incompatibility inducing symbionts may have negative effects in mass-rearing and field applications. The symbionts living in the digestive tracts of the beneficials may be of special interest as many beneficials are reared on artificial diets that contain antimicrobial preservatives. Biological control in general will benefit from a better understanding of the symbionts involved in both pests species and their natural enemies. Unfortunately these subject receive only limited attention leaving the potential benefits unexplored.

/	)
1	
$\vdash$ $-$	 
1	
1	
1	
1	
<u> </u>	 
<u> </u>	 
$\vdash$ $-$	 
1	
1	
1	
$\vdash$ $-$	 
1	
1	
1	
1	
1	
1	
1	
1	
1	 i a
1	
1	
1	
1	
1	
1	
\	/
\	/
\	
	/

## Regulatory aspects and safety considerations in the EU

Aguilera J., Gomes A.R. and Olaru I.

European Food Safety Authority. Via Carlo Magno 1/a, 43126 Parma, Italy - Jaime.Aguilera@efsa.europa.eu

Paratransgenesis, or the genetic modification of microbial symbionts to confer new traits to their hosts, has been identified as a promising tool to fight arthropods which are pests or vectors of infectious diseases. The applied use of paratransgenesis would involve the deliberate release into the environment of genetically modified micro-organisms (GMMs) or arthropods carrying them. thereby raising questions about health and environmental safety. The European Union has different legislative instruments in place to ensure the safe use of pesticides and insecticides (including those of microbial origin) as well as GMMs. Depending on their purpose, paratransgenic insects or their GM symbionts may fall under one or more of those legislations. A common requirement to be fulfilled by those who intend to commercialise such products is to conduct a scientific risk assessment to determine whether the product is safe for humans. animals and the environment. The European Food Safety Authority (EFSA) has a key role in the evaluation of applications for marketing genetically modified organisms and plant pesticides, and therefore would assess the safety of GMMs to be used as plant pest controllers via paratransgenesis. EFSA has published guidance which explains the strategy and the scientific data necessary to conduct the risk assessment of GMMs. Future EFSA guidance on the risk assessment of GM insects will provide elements that may also be applicable for paratransgenic insects. EFSA opinions are taken into account by the different European regulatory authorities prior to a decision regarding authorisation to commercialise the product.

Keywords: Paratransgenesis, GMM, biocontrol, biopesticide, risk assessment

/	)
<u> </u>	 
1	
$\vdash$ $-$	 
1	
1	
1	
1	
<u> </u>	 
<u> </u>	 
$\vdash$ $-$	 
1	
1	
1	
$\vdash$ $-$	 
1	
1	
1	
1	
1	
1	
1	
1	
1	 i a
1	
1	
1	
1	
1	
1	
\	/
\	/
\	
	/

### Mites, symbionts and commercial implications

### Perotti M.A.

School of Biological Sciences, University of Reading, Reading, RG6 6AS, UK. - m.a.perotti@reading.ac.uk

Acarine Biocontrol Agents or ABAs are of extraordinary importance for the control of pest and vectors. ABAs are mass-produced and commercialised worldwide. Good examples are mites of the family Phytoseiidae, which have become regular components of agricultural systems where, as predators, they assist with suppressing pest mite/insect populations that affect crops.

ABAs, like many other arthropods, also engage in a variety of symbiotic associations with microorganisms that induce a wide diversity of phenotypes. For example, mite-bacteria interactions can be manifold and may involve manipulations of the mites rearing conditions to enhance productivity. Therefore, symbionts are not longer exclusive to symbiont-based pest control strategies because they can also be used to manipulate and increase the production of beneficial arthropods. The use of these mite-microbial interactions is leading to a new way of commercialization of ABAs: All-in-one ABAs, where the beneficial hosts (ABAs) are produced following new symbiont-based enhanced production systems.

Commercialization procedures must be reconsidered. New rules for accessing, patenting, exporting, importing and/or releasing All-in-one ABAs are required and these must follow rules of utilization of these genetic resources in accordance with international conventions and protocols.

Keywords: acari, mites, acarology, biological control, pest control

	/																													
1																														
	-	_	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	-
											_														_					
	-	_	_	_	_	_	_	_	_		_	_	_	_	_	_	_	_	_	_	_	_	_	_		_	_	_	_	_
	-	_	_	_	_	—	_	_	_	_	—	—	_	_	_	—	_	_	_	_	—	_	_	_	—	—	—	_	_	_
	-		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	
			_	_	_		_	_	_			_	_	_		_	_	_	_		_	_	_	_		_	_	_	_	
	-	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_
-	-	_	—	—	—	—	—	—	_	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	_	_	-
	-	_	—	_	—	—	—	—	_	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	_	-
			_	_	_	_	-	_		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_			_	_	_	
1																														
		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_
$\vdash$	-	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	-
-	-	—	_	_			—	—	-	—	—	—	—	_	—	_	—	—	—	—	—	—	—	-	—	—	—	—	—	-
1																														
	-	_	_	_		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	-	-0	2
																												100		
																												~		
																												/		
																												1		
																										~!				
																								~	~	~				
																														1

### Is it mine? Ethical and other considerations regarding the manipulation of the (human) microbiome or (human) symbionts

Braig H. R.

### School of Biological Sciences, Bangor University, Bangor, Wales, UK; h.braig@bangor.ac.uk

Humans have 10-times more bacterial than human cells accounting for roughly 100-times more bacterial genes than human genes, which suggests that the human microbiome might contribute significantly to the definition of a human self. These bacteria can be found on the skin, in the mouth, nose, esophagus, digestive tract and vagina. Such association is neither unique to humans as it applies to arthropods as well, nor is its discovery new. In 1942, Edward A Steinhaus already produced a Catalog of Bacteria Associated Extracellularly with Insects and Ticks. And it is not only bacteria; it can be archaea, fungi, parabasalids, heterokonts, even mites. Some of the associations are very short-lived and much influenced by the environment leading to an ephemeral bacterial fingerprint while others constitute a core microbiome.

Ethics or moral philosophy is the basis from which the general legal system has evolved in a very slow evolutionary process driven by thinkers and the populace. However, matching the processes characteristic of a punctuated equilibrium, the ethics and law of dental hygiene, the ethics and law of genomes and environmental regulation, just to name a few examples, have rather suddenly arisen. This no longer seems to be driven by the populace but by a few academics and a few associations, examples being the International Society for Environmental Ethics, The Animal Welfare Science, Ethics & Law Veterinary Association, and so on. This raises the questions of who is leading the discussion on the ethics of microbiomes and symbionts.

Issues concerning microbial transplantation, pharmacomicobiomics and microbial profiling are most prominent. Ethical questions of privacy and confidentiality, consent, ownership and governance emerge. Microbial profiling might not only reveal information about diet (vegetarian or not) or shape (fat or slim) of an individual, it also might hold information about predisposition for caries, inflammatory bowel disease, Crohn's disease, asthma, type 1 diabetes, rheumatoid arthritis; life expectancy, cultural identity, caste or social characteristics. For insects and mites, the microbiome might determine fitness, the ability to vector diseases as important as dengue and malaria, and the ability to mass rear insects for sterile insect release or as predatory mites for biological control.

Ethical issues of bioprospecting (biopiracy) and return of results are interrogated from the point of patentability of microbial associations. Using caries-preventing Streptococcus bacteria and insect-symbiont associations as examples, the current legal interpretation making naturally occurring bacteria even in new combination unpatentable and returnless for its discoverer is questioned. Lawmakers and judges in the last century could not have imagined the impact natural symbiotic associations might have on human health and economic productivity. What in the past has been considered as dust, debris, dirt or waste and feces, has become an important and defining commodity. The question whether or not sequence variations in natural genes are patentable is still open, but should this evolve as being the case, variations in naturally occurring symbionts might be the next challenge; in terms of legal interpretation, a very slippery slope. Would a change in law benefit the research community?

Keywords : ethics, ownership, legal protection, beneficial bacterial, naturally occurring bacteria

/	)
<u> </u>	 
1	
$\vdash$ $-$	 
1	
1	
1	
1	
<u> </u>	 
<u> </u>	 
$\vdash$ $-$	 
1	
1	
1	
$\vdash$ $-$	 
1	
1	
1	
1	
1	
1	
1	
1	
1	 i a
1	
1	
1	
1	
1	
1	
\	/
\	/
\	
	/

### Research on arthrophod symbioses in Latvia

Jankevica L.¹, Metla Z.¹ and Bormane A.²

¹Institute of Biology, University of Latvia, Miera street 3, Salaspils, LV - 2169, e-mail: jankevica.liga@inbox.lv, ²Centre of Disease Prophylaxis and Control, Klijanu street 7, Riga, LV 1012, - antra.bormane@gmail. com

Research on biological control has been carried out in the Institute of Biology since 1967. The Laboratory of experimental entomology deals with all basic and practical aspects of biological control. The main attention is paid to ecology of pest organisms and their pathogens (viruses, bacteria and fungi). In recent years microflora of pollinator honey bee (*Apis mellifera*) and different forest pests (Lepidoptera, Hymenoptera) was investigated. The impact of different biotic and abiotic factors on associations of microorganisms was observed. Our interest is based on symbiotic control using associated microorganisms. The laboratory studies interactions between insect pathogenic bacteria and bacteria, that belong to the genera *Bacillus* and *Lactobacillus*. The efficiency of the active bacteria is also being tested.

Research on vectors has been going on for more than twenty years in Latvia. Tick-borne encephalitis (TBE) and Lyme borreliosis (LB), the latter caused by spirochaete bacteria vectored by the same tick species, are significant vectorborne diseases in Latvia. Abundance and sesonal dynamics of ticks *Ixodes ricinus* and *Ixodes* persulcatus and infection prevalence in tick populations with pathogenic agents were regulary monitored by scientists from Centre of Disease Prophylaxis and Control. There are registred clinical cases of borreliosis and cases of erlihiosis. In reecent years infections in tick populations with *Borrelia burgdorferi* was in average 30 %; with *Anaplasma phagocitophylum* - in 5%, TBE infection (varies by the year) in range 2-7%. In future attention will be paid to elaboration of new methods of diagnostic and control of nonviral tick-born infections.

Keywords: associations of microorganisms, ticks, vectors

	/																													
1																														
	-	_	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	-
											_														_					
	-	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_		_	_	_	_	_
	-	_	_	_	_	—	—	_	_	_	—	—	_	_	_	—	_	_	_	_	—	_	_	_	—	—	—	_	_	_
	-		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	
			_	_	_		_	_	_			_	_	_		_	_	_	_		_	_	_	_		_	_	_	_	
	-	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_
-	-	_	—	—	—	—	—	—	_	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	_	_	-
	-	_	—	_	—	—	—	—	_	—	—	—	_	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	_	-
			_	_	_	_	-	_		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_			_	_	_	
1																														
		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_
$\vdash$	-	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	—	_	_	_	-
-	-	—	_	_			—	—	-	—	—	—	—	_	—	_	—	—	—	—	—	—	—	-	—	—	—	—	—	-
1																														
	-	_	_	_		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	-	-0	2
																												100		
																												~		
																												/		
																												1		
																										~!				
																								~	~					
																														1

### Dynamic Wolbachia infection of Acromyrmex leafcutter ants: potential for a nutritional symbiosis

Andersen S.B.¹, Boye M.², Nash D.R¹. and Boomsma J.J.¹

¹Centre for Social Evolution, University of Copenhagen, Universitetsparken 15, 2100 Copenhagen, Denmark SBAndersen@bio. ku.dk ²National Veterinary Institute, Technical University of Denmark, Bülowsvej 27, DK-1790 Copenhagen, Denmark

Wolbachia are renowned as reproductive parasites, but their phenotypic effects in eusocial insects are not well understood. We used a combination of grt-PCR, fluorescence in situ hybridisation, and laser scanning confocal microscopy to evaluate the dynamics of Wolbachia infections in the leafcutting ant Acromyrmex octospinosus across developmental stages of sterile workers. We confirm that workers are infected with one or two widespread wsp genotypes of *Wolbachia*, show that colony prevalences are always 100%, and characterize two rare recombinant genotypes. One dominant genotype is always present and most abundant while another only proliferates in adult workers of some colonies and is barely detectable in larvae and pupae. An explanation may be that Wolbachia genotypes compete for host resources in immature stages while adult tissues provide substantially more niche space. Tissue-specific prevalences of the two genotypes differ, with the rarer genotype being overrepresented in the adult foregut and thoracic muscles. Both genotypes occur extracellularly in the foregut, suggesting an unknown mutualistic function in worker ant nutrition. Both genotypes are also abundant in the faecal fluid of the ants, suggesting that they may have extended functional phenotypes in the fungus garden that the ants manure with their own faeces.

Keywords : esocial ant host, extracellular Wolbachia, FISH, nutritional symbiosis, conflict

## The microbiome of the pest mite Rhizoglyphus robini: implications for the pest's status

Zindel R.¹, Ofek M.², Minz D.², Palevsky E.³, Zchori-Fein E.³ and Aebi A.¹

¹Agroscope Reckenholz-Tänikon, Research Station ART, Reckenholzstr. 191, 8046 Zürich, Switzerland; ²Institute of Soil, Water and Environmental Sciences, Agricultural Research Organization, Bet Dagan, Israel. ³Department of Entomology, Newe Ya'ar Research Center, Agricultural Research Organization, Ramat Yishay, Israel - renate. zindel@art.admin.ch

The biology of many arthropods can only be understood when the associated microbiome is also considered. The bulb mite *Rhizoglyphus robini* Claparede (Acari: Astigmata: Acaridae), a pest on Liliacean cultures, has previously been observed on fungus infected plant cultures and preferred infected plants over uninfected onion seedlings in choice tests. We investigated the composition of its microbial associates and were able to link it to its nutritional preferences. We described the bacterial community using 454 sequencing of 16S rRNA gene fragments and DGGE, finding three genera to be dominant: *Myroides* (41.4% of total sequences), *Serratia* (11.4%) and *Alcaligenes* (4.5%), of which *Serratia* and *Alcaligenes* are known chitinase producers. We showed that mite fecundity was indeed higher when feeding on fungi and that chitin was digested by mite homogenate. Bacteria were shown to be responsible for chitinase production. We conclude that *R. robini*, at least to a certain extent uses fungal chitin as a food source with the help of its associated bacteria. This knowledge on the mite's biology supports the general concept of multi genome organisms.

Keywords: bacterial symbionts, chitin digestion, 16S rRNA pyrosequencing, DGGE

## The genome sequence of *Midichloria mitochondrii* and the origin of mitochondria

Sassera D.¹, Lo N.², Epis S.², D'Auria G.³, Montagna M.¹, Comandatore F.¹, Horner D.¹, Peretó J.³, Luciano AM.¹, Franciosi F.¹, Ferri E.⁴, Crotti E.¹, Bazzocchi C.¹, Daffonchio D.¹, Sacchi L.⁵, Moya A.³, Latorre A.³ and Bandi C.¹

¹Università degli Studi di Milano. Via Celoria 10, Milano, Italy; ²University of Sydney, Australia^{; 3}Universidad de Valencia, Spain^{; 4} Università di Milano Bicocca, Italy^{; 5}Università degli Studi di Pavia, Italy - davide. sassera@unimi.it

The sheep tick *lxodes ricinus*, an arthropod of high medical and veterinary importance, harbours a peculiar intracellular symbiont, the alphaproteobacterium *Midichloria mitochondrii*. This symbiont is a member of the order *Rickettsia*les, bacteria which are considered, based on phylogenetic analyses, to be the closest extant relatives of mitochondria. Furthermore, *M. mitochondrii* is not only phylogenetically close to the mitochondrion, it also presents a unique type of interaction with it. *M. mitochondrii* is in fact able to invade the mitochondria of the ovary cells of its tick host, specifically localizing in the intramembrane space, between the inner and outer membranes of the organelles.

We decided to perform whole genome sequencing of *M. mitochondrii* in order to shed light on this unique symbiosis. We developed a specific method, consisting of single oocyte isolation by micromanipulation followed by multiple displacement amplification and sequencing with 454 and Sanger technologies. The genome was assembled using Mira assembler, Bambus scaffolder, Gap4 for contig viewing and ad-hoc bioinformatics tools for finishing. Inverse-PCRs were performed to close gaps and disambiguate repetitive regions.

Genome annotation revealed the presence of genes coding for a complete flagellar apparatus and for the *cbb3* cytochrome oxidase, both previously unreported in *Rickettsia*les. Phylogenomics analyses showed these genes to be vertically inherited from the common ancestor of *Rickettsia*les and mitochondria. These findings suggest a novel scenario on the origin of eukaryotes, with the ancestor of mitochondria playing a more active, parasitic role, in a microaerophilic habitat.

Keywords: Ixodes ricinus, Rickettsiales, mitochondria, eukaryogenesis, flagellum

#### On tsetse flies-Wolbachia symbiotic associations

Doudoumis V.¹, Tsiamis G.¹, Wamwiri F.^{2,6}, Brelsfoard C.^{2,3}, Alam U.², Aksoy E.², Dalaperas S.¹, Egyir-Yawson A.⁴, Malele I.⁵, Ouma J.⁶, Takac P.⁷, Abd-Alla A.⁸, Aksoy S.² and Bourtzis K.^{1,9}

¹Department of Environmental and Natural Resources Management, University of Western Greece, 2 Seferi St, 30100 Agrinio, Greece; ²Yale University School of Public Health, 60 College St., 811 LEPH, New Haven, CT 06520 USA; ³Current address: Department of Entomology, University of Kentucky, S-225 Ag. Science Center North, Lexington, KY 40546 USA; ⁴Vector Genetics Laboratory, Department of Animal Science, Biotechnology & Nuclear Agriculture Research Institute, Ghana Atomic Energy Commission, P.O. Box LG 80, Legon, Accra, Ghana; ⁵Tsetse & Trypanosomiasis Research Institute (TTRI), Majani Mapana, Off Korogwe Road, P. O. Box 1026 Tanga, Tanzania; ⁶Trypanosomiasis Research Centre, Kenya Agricultural Research Institute, P.O. Box 362, Kikuyu 00902 Kenya; ⁷Institute of Zoology, Department of Entomology, Slovak Academy of Science, Dubravska cesta 9, 845 06 Bratislava, Slovakia; ⁸Insect Pest Control Laboratory, Joint FAO/IAEA Division of Nuclear Techniques in Food and Agriculture, Vienna, Austria; 'Biomedical Sciences Research Center Al. Fleming, 16672 Vari, Greece - edoudoum@cc.uoi.gr

Wolbachia is a group of endosymbiotic  $\alpha$ -Proteobacteria which infect numerous arthropod species inducing reproductive abnormalities such as cytoplasmic incompatibility (CI), parthenogenesis, feminization and male killing. Because of its unique properties, Wolbachia is currently being considered as a promising tool for the biological control of arthropod pests and disease vectors including *Glossina* spp. In the present study, we employed a Wolbachia specific 16S rRNA gene PCR assay to investigate the presence of Wolbachia in a total of 5339 specimens of eleven different Glossina species, originating from laboratory stocks and natural populations from thirteen African countries. The genotyping of the Glossina Wolbachia strains was based on the Multi Locus System Typing (MLST), the wsp and 16S rRNA genes. During this analysis, it was shown that fragments of at least six Wolbachia genes (16S rRNA, fbpA, ftsZ, wsp, gatB and coxA) have been transferred into tsetse fly chromosomes and have been pseudogenized by extensive deletion events. The complete characterization of the multiple Wolbachia genomes, cytoplasmic and nuclear, present in tsetse flies will shed light into the host-Wolbachia symbiotic associations and will contribute to the development and use of effective Wolbachia-based strategies for the control of tsetse flies and trypanosomosis.

Keywords: Wolbachia, Glossina, horizontal gene transfer

#### Detection of spiders microbial communities in Madeira Island

Moreira M.¹, Santos D.², Latorre A.² and Khadem M.¹

¹ISOPlexis Banco de Germoplasma, Universidade da Madeira, Funchal 9000-390 Portugal; ²Institut Cavanilles de Biodiversitat i Biologia Evolutiva, Universitat de Valencia, P.O. Box 22085, Valencia, Spain marta_amoreira@hotmail.com

Spiders harbour heritable bacteria that are known to be reproductive manipulators in arthropods. However, only few studies have focused on spiders' microbial communities and the role that they may play in their biology. We have investigated the endoymbionts of spiders from oceanic island of Madeira, home to many endemic species of spiders. A total of six samples from four different species, Pholcus phalangioides, Cyrtophora citricola and Segestria florentina and the endemic one Hogna insularum, were screened for the presence of endosymbionts using universal 16S rDNA primers. Three samples were positive: two from P. phalangioides and one from H. insularum. The positive samples were amplified and sequenced for three gene fragments, 16S rDNA, ATP synthase F1 alpha subunit (*atpA*) and subunit I of cytochrome C oxidase (*coxA*). Multiple infections by Rickettsia and Wolbachia were found in both samples of P. phalangioides, while only one type of bacteria was present in H. insularun. Phylogenetic analysis of 16S rDNA gene sequences clustered with the Rickettsia from basal ancestral group of Limoniae, together with those from water beetles (Deronectes) and predatory bugs (Macrolophus). Wolbachia sequences of the16S rDNA gene were clustered with those reported in supergroup A. H. insularum was infected by Sphingomonas bacteria presumably due to other contamination. The quality of sequences for other genes was not high enough for phylogenetic analysis.

Keywords: Spiders, Endosymbionts, Rickettsia, Wolbachia, Madeira Island

# Effect of high temperature on distribution of pathogenic *Wolbachia* strain wMelPop in *Drosophila* melanogaster brain cells

Strunov A.¹, Kiseleva E.¹ and Gottlieb Y.²

¹The Institute of Cytology and Genetics, Prospekt Lavrentyeva 10, Novosibirsk, 630090, Russian Federation; ²The Koret School of Veterinary Medicine, Faculty of Agriculture, POB 12, Rehovot, 76100, Israel - strunov.anton@gmail.com

Symbiotic bacteria together with their host are exposed to various environmental stresses. High temperature is one of the stressful factors that have a strong impact on organism biology. In *Wolbachia-Drosophila* association elevated temperature has diverse effects depending on the host species and bacterial strain. *Wolbachia* strain wMelPop actively proliferates in D. melanogaster brain and muscle tissues at 29°C and strongly reduces the longevity of its host (Min&Benzer, 1997). Due to the potential use of this pathogenic strain as a bio-control agent the purpose of my STSM project was to investigate the effect of high temperature (29°C) on wMelPop distribution in *D. melanogaster* larva and adult brain using FISH and confocal microscopy.

The data obtained has shown a strong heterogeneity of bacterial distribution in larva as well as in adult brain. There were cells free from *Wolbachia* and cells fully packed with them. Surprisingly,  $29^{\circ}$ C did not affect the bacterial distribution in  $3^{rd}$ -instar larva brain. *Wolbachia* titer and localization were similar to  $25^{\circ}$ C. However, the bacterial titer significantly increased already at the first day of imago stage at  $29^{\circ}$ C and then continued to grow. Bacteria dramatically over-replicated and filled almost each area of adult fly brain by  $13^{th}$  day of imago stage at high temperature. In contrast with that the bacterial titer at  $25^{\circ}$ C didn't changed a lot by this day. What has happened with *Wolbachia* during the pupa stage? The question will be discussed.

Keywords: Wolbachia, Drosophila melanogaster, brain, high temperature, FISH

#### Manipulation of mating behavior

<u>Schneider D.I.</u>¹, Chao T.², Ehrman L.², Däuble W.¹, Kaltenpoth M.³ and Miller W.J.¹

¹Lab of Genome Dynamics, Dep. Cell and Developmental Biology, Medical University Vienna, Austria; ²State University of New York, Division of Natural and Social Sciences, Purchase, USA; ³Research Group Insect Symbiosis, Max Planck Institute for Chemical Ecology, Jena, Germany. daniela.schneider@meduniwien.ac.at

Recent research focuses on biosynthesis, expression, and biological importance of sex pheromones in insects. From these studies, we have gained insight into how crucial these complex chemical compounds are for inter-individual communication, including recognizing mating partners. Mate recognition in turn, is highly significant for sexual isolation, a key parameter of speciation. Like many other insects, *Drosophila* use a cocktail of long-chain hydrocarbons (CHCs) as sex pheromones. The composition of cuticular hydrocarbons, i.e., the pheromone pattern of an individual is characteristic, and recent studies have demonstrated that quantitative alterations in these profiles contribute significantly to sexual isolation in insects.

Extraction, detection, and precise quantification of CHCs, some occurring at quite low concentrations, are often impeded by availability and accessibility of the tested specimen, as well as the detection method itself. Hence, highly sensitive and reliable methods are needed for such analyses. We have taken advantage of the excellent equipment and expertise provided by the Research Group Insect Symbiosis at the Max Planck Institute for Chemical Ecology in Jena, Germany. This group has recently reported on a technically improved method for unambiguous and quick determination of CHCs in insects (Kroiss et al. 2011). By applying this GC-MS technique to the *D. paulistorum-Wolbachia* model system, we were able to detect and compare CHC patterns between wildtype and *Wolbachia*-knockdown lines. The improved method allowed for significant reduction of sample size than the one used before. Further, a more exact analysis of the major compounds could be performed.

Keywords: Drosophila paulistorum, cuticular hydrocarbons, gas chromatography-mass spectrometry

## Role of bacterial endosymbionts for the use of predatory bugs in biological pest control

#### Machtelinckx T., Van Leeuwen T., Gheysen G. and De Clercq P.

Laboratory of Agrozoology, Department of Crop Protection, Coupure Links 653, B-9000 Gent, Belgium. Thijs.machtelinckx@ugent.be

Symbiotic micro-organisms are widespread in arthropods in which they play an important evolutionary and ecological role. Although such endosymbionts are well-studied, their possible use in biological control programmes is still underestimated, and generally focuses on their potential effect in pest insects. However, as these endosymbionts are able to manipulate the reproduction of their arthropod host, their presence in beneficial insects may also have a substantial impact on the use of such insects in biological control. Whereas in the past the focus of endosymbiont research in beneficial insects was most often on parasitoids, this paper will illustrate the importance of endosymbionts in predatory bugs.

The presence and role of endosymbionts in predators has been best studied in ladybugs, which are known to be infected with male killing inducing endosymbionts. Besides coccinellids, heteropteran predators have proven to be excellent biological control agents, and their zoophytophagous feeding habits make them an interesting case study to explore the importance of symbiosis in insects possessing this feeding habit. Research on the economically important predator *Macrolophus pygmaeus* indicated that endosymbionts induce a strong cytoplasmic incompatibility effect in its host. Furthermore, the wide variance in infection frequencies between different *M. pygmaeus* populations may have a substantial impact on the predator's population growth, and as such negatively influence biological control programmes employing this predatory mirid either at the production level or in the field.

These findings demonstrate that it would be warranted to expand research on endosymbiotic bacteria to other heteropteran predators in order to elucidate their potential role on the development, reproduction and feeding ecology of this group of economically important natural enemies.

Keywords: Predatory bugs, Heteroptera, Coccinellidae, endosymbionts

																												$\backslash$
/																												
																												1
	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	-
	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	
L _	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_
<u> </u>	_	—	—	_	_	—	_	—	—	—	—	—	—	—	—	—	_	—	_	—	—	_	—	—	—	_	_	_
$\vdash$ $-$	_	_	—		—	_	—	_	—	—	_	_	—	—	—	—	—	—	—	—	—	_	—	—	_	—	—	$\neg$
1																												
1																												
1																												
1																												
1																												
	-	_	_	_	_	_	_	_	—	_	_	_	_	_	_	_	_	_		_	_	_	_	_	_	_	_	
		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	
	_	_	_		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_
<u> </u>	_	_	_			_	_	_	_	—	_	_	_	—	_	_	_	_	—	_	_	_	_	_	_	_	_	-
	_	—	—	_	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	-
	_	_	_		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_
1																												
1																												
1																												
1																												
		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	
	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_		_	_	_	_	_	
1																												
1																												
L _	_		_	_	_	_		_	_	_	_		_	_				_		_	_	_	_	_	_	_	_	
								-	-	-		_		_		-				_	_	_	_	-		_	_	
1																												
1																												
1																												
1																												
						_		_	_	_	_	_	_	_	_	_				_	_		_		_	_	_	
1																												
1																												
1																												
1																												
1																												
			_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	-		_
																											Â	·
																										1		
																										X		
																									11			
																									$\mathbb{Z}$			
1																								1		/		
1																						~	~	$\sim$				
																						-		-				
\																												
																												/
$\sim$																											/	

### Posters

## Genomic basis and possible evolutionary consequences of ectosymbiotic lifestyle in a sulfide-rich environment

Cerveau N., Bauermeister J. and Dattagupta S.

Georg-August-Universität Göttingen, Courant Research Center Geobiology, Goldmischmidstraße 3, 37077 Göttingen, Germany nicolas.cerveau@geo.uni-goettingen.de

A broad continuum exists in the level of integration between hosts and symbionts. Ectosymbioses have been suggested to be the first step towards the evolution of more integrated extra- and intracellular symbioses, leading eventually to the development of obligate endosymbionts or organelles. From this perspective, ectosymbioses offer the best opportunity to study the early stages of the establishment of symbioses.

Ectosymbioses are common in sulfur-rich marine environments such as deep-sea hydrothermal vents, cold seeps, and organic-rich coastal sediments. Invertebrates living in these habitats are exposed to diverse free-living bacterial communities and are susceptible to colonization by many opportunistic, non-specific surface-dwellers. In this kind of habitat, invertebrate have developed stable ectosymbiotic associations with some epsilon and gammaproteobacteria. It has long been documented that *Thiothrix*, a gammaproteobacteria belonging to the Thiotrichaceae family are ectosymbionts of marine amphipod. Recently, *Thiothrix* bacteria were also identified as ectosymbionts of the freshwater amphipod *Niphargus* ictus from the Frasassi caves in central Italy. These caves harbor various sulfide-oxidizing bacteria, notably belonging to the *Thiothrix* genus, which form bacterial mats. *Thriothrix* bacteria are abundant in mats but the free-living strains are distinct from the bacteria found in association with N. ictus.

Our aim is to use this atypical association between *Thriothrix* and N. ictus in a fresh water ecosystem to investigate the early phase of the symbiosis. The symbiont genome will be compared with that of free-living *Thiothrix* present in bacterial mats in the amphipod habitat to uncover the genomic basis and possible evolutionary consequences of symbiotic lifestyle.

Keywords: ectosymbiont, genomic variation, sulfide-rich environment

#### New Supergroups and hidden Wolbachia diversity in aphids

Augustinos A.A.^{1,9}, Santos-Garcia D.², Dionyssopoulou E.^{1,9}, Moreira M.³, Papapanagiotou A.⁴, Scarvelakis M.¹, <u>Doudoumis V.^{1,9}</u>, Ramos S.², Aguiar A.F.⁵, Borges P.A.V.⁶, Khadem M.³, Latorre A.^{2,7}, Tsiamis G.^{1,9} and Bourtzis K.^{1,8,9}

¹Department of Environmental and Natural Resources Management, University of Ioannina, 2 Seferi St., Agrinio 30100, Greece; ²Institut Cavanilles de Biodiversitat i Biologia Evolutiva, Universitat de València, Apartado Postal 2085, 46071 Valencia, Spain; ³ISOPlexis Gene Bank, Universidade da Madeira, Funchal 9000 Portugal; ⁴Department of Greenhouse Crops and Floriculture, Technological Educational Institute of Messolonghi, Greece; ⁵Laboratório de Qualidade Agrícola, Núcleo de Fitopatologia, Camacha 9135-372, Madeira, Portugal ⁶Universidade dos Acores, Departamento de Ciências Agrárias CITA-A (Azorean Biodiversity Group), 9700-042 - Angra do Heroísmo, Terceira -Azores; ⁷Área de Genómica y Salud, Centro Superior de Investigación en Salud Pública (CSISP), Avenida de Cataluña 21, 46020 Valencia, Spain; ⁸Biomedical Sciences Research Center Al. Fleming, 16672 Vari, Greece; ⁹Present address: Department of Environmental and Natural Resources Management, University of Western Greece, 2 Seferi St., Agrinio 30100, Greece - edoudoum@cc.uoi.gr

Aphid species have a major impact on agriculture since they are significant pests and also act as vectors of numerous plant diseases. They are easily dispersed by wind and due to the parthenogenetic mode of reproduction small populations can become established and rapidly increase in number. Therefore aphids are considered as significant economically invasive pests throughout the world. The about 4000 species worldwide engage in highly interesting and complex relationships with their microbial fauna. One of the key symbionts in arthropods is *Wolbachia*, an  $\alpha$ -Proteobacterium implicated in many important biological processes and believed to be a potential tool for biological control. Aphids were thought not to harbour Wolbachia; however, current data suggest that its presence in aphids has been missed. The aim of the present study is to map the Wolbachia infection status of natural aphid populations, along with the characterization of the detected Wolbachia strains. Out of 426 samples from Spain, Portugal, Greece, Israel and Iran, 37 were found to harbour Wolbachia. Our 16S rRNA based data indicate the presence of two new Wolbachia supergroups in aphids, along with some strains belonging either to supergroup B or to supergroup A. Further characterization of the new aphid supergroups could not be completed using MLST and/or sequencing data from the groEl, gltA and wsp genes. This could be attributed to differences in the titre of the infection, multiple infections, and/or high divergence of the newly described Wolbachia strains of aphids. New tools are needed to further analyse the aphids Wolbachia strains.

Keywords: Wolbachia, insect symbiosis, aphids, 16S rRNA

#### Bacterial symbionts of Melophagus ovinus (Diptera: Hippoboscidae): a comparative model to the symbiotic system in tsetse flies

Husník F., Nováková E., Chrudimský T. and Hypša V.

Faculty of Science, University of South Bohemia & Institute of Parasitology, Biology Centre of ASCR, Branišovská 31, České Budějovice 37005, Czech Republic

Filip Husník: filip@paru.cas.cz, Eva Nováková: novaeva@paru.cas.cz, Tomáš Chrudimský: tomtom@paru.cas.cz, Václav Hypša: vacatko@paru. cas.cz

Insects living exclusively on vertebrate blood, such as lice, bedbugs or tsetse flies, rely on symbiotic bacteria to provide them with nutrients missing from their blood diet (mainly B-vitamins and cofactors). The sheep ked Melophagus ovinus (Diptera: Hippoboscidae) is a wingless obligatory blood-sucking parasite of sheep with adenotrophic viviparity and nourishment of larvae through milk glands. Here, we present basic morphological and molecular characteristics of its symbiotic system and compare it with the best known symbiotic system in tsetse fly (Diptera: Glossinidae). Similar to the tsetse flies, the sheep keds possess a specialized section of midgut (bacteriome) which harbors highly adapted obligatory mutualists transmitted vertically through the milk glands. Unlike the tsetse symbionts of the genus Wigglesworthia, these bacteria belong to the large and biologically diverse genus Arsenophonus (Gammaproteobacteria: Enterobacteriales). In analogy to the tsetse inhabiting symbiotic system, this symbiont is accompanied by a facultative symbiotic bacterium infecting various tissues, which originated within Sodalis lineage (Gammaproteobacteria: Enterobacteriales) independently of Sodalis glossinidius. Along microvilii of the midgut, two additional microorganisms are commonly present: Bartonella melophagi (Alphaproteobacteria: Rhizobiales) and Trypanosoma melophagium (Excavata: Kinetoplastida). Our results show that Melophagus ovinus contains functionally convergent, but phylogenetically independent symbiotic system to that in tsetse fly and can provide a useful comparative model of symbiosis within blood-sucking insects.

Keywords: hippoboscid fly, Arsenophonus, tsetse, Sodalis, Bartonella

#### Endosymbiotic Archaea - Yes, they do exist!

#### Lind A.E.¹ and Ettema T.J.G.²

¹Laboratories of Genome Dynamics, Centre of Anatomy and Cell Biology, Medical University of Vienna, Austria. E-mail: anders.lind@meduniwien. ac.at. ²Department of Cell and Molecular Biology; Molecular Evolution, Biomedical Center, Science for Life Laboratory, Uppsala University, Uppsala, Sweden - thijs.ettema@icm.uu.se

While there are many organisms that have adapted to an intracellular lifestyle, whether as mutualists, commensals or parasites, almost all come from the same domain of life, bacteria. Even now, some 20 years after the first discovery of intracellular Archaea, few studies are being done on these organisms.

One such Archaea can be found within the eukaryotic ciliate *Nyctotherus ovalis*, whom itself is part of the hindgut micro fauna of several cockroach species. While there are other protists that harbour these kinds of endosymbionts, N. ovalis has some additional remarkable features. The hydrogen producing hydrogenosomes of *N. ovalis* contains the unique feature of having maintained a genome, whereas hydrogenosomes from other species have lost their genetic material over time.

The symbiotic Archaea found in *N. ovalis* are methanogens, converting the hydrogen from the hydrogenosomes into methane, thus providing the host with the benefit of lowering the partial pressure of hydrogen.

Because of the endosymbiotic lifestyle of these archaeons, they cannot be readily grown in pure culture. This has resulted in a lack of sequence data from these organisms. We have taken a single cell approach to sequencing of the complete genome of these organisms. Single cells were sorted and multiple displacement amplification was used in order to obtain enough genomic material for Illumina sequencing. The data obtained has been used to get a better understanding of these organisms, and to further establish the phylogeny of these Archaea.

Keywords: Archaea, Methanobrevibacter, endosymbiont, Nyctotherus ovalis

																											~	
																												$\backslash$
(																												
																												1
	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	-
																							_					
<u> </u>		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_
	—	_	_	—	_	_	_	_	_	_	_	_	_	—	_	_	_	_	—	_	_	_	—	—	—	_	_	_
	_	_	_	_	_	_	_	_		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	
	_	_	_	_	_	_	_	_		_	_	_		_	_	_	_		_	_	_	_		_	_	_	_	
		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_
<u> </u>	_	—	—	—	—	—	_	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	_	—	_
	_	—	—	—	—	—	—	—	—	—	_	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	-
		_	_	_	_	_	_	_			_	_	_	_	_	_	_	_	_	_	_	_		_	_	_	_	
	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_
	_	_	-	-	-	-	-	-	-	_	_	_	_	_	_	-	-	-	-	-	-	-	-	-	-	-		1
	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_		_	_	_	_	_
$\vdash$ $-$	—	_	—	—	—	—	_	_	—	—	—	_	—	—	—	—	_	—	—	—	—	_	—	—	—	—	—	$\neg$
		—	_	—	—	—		—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	$\neg$
													_	_	_		_									-		
		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	6	7
																										100		
																										5		
																										/		
																										1		
1																							~ ~	~!				
																						~	~					
																												1

## Evolutionary stasis in Blattabacterium, an ancient primary endosymbiont

Patiño-Navarrete R.¹, Ponce De León M.², Montero F.², Moya A.^{1,3,4}, Peretó J.^{1,5} and Latorre A.^{1,3,4}

¹Institut Cavanilles de Biodiversitat i Biologia Evolutiva, Universitat de València, P.O. Box 22085, E-46071 València, Spain. ²Departamento de Bioquímica y Biología Molecular I, Facultad de Química, Universidad Complutense de Madrid, Spain. ³Departament de Genètica, Universitat de València. ⁴Centre for Public Health Research (CSISP), València. ⁵Departament de Bioquímica i Biologia Molecular, Universitat de València - pereto@uv.es

Many insect species have established long-term symbiotic relationships with intracellular bacteria. Among these associations, the case of the cockroaches and Blattabacterium cuenoti is one of the most ancient mutualistic symbioses described up today. In the present work we present the complete genome sequence of the B. cuenoti symbiotic strain from the cockroach Blatta orientalis (BBor). The comparison of this genome with those previously published from the cockroaches Blattella germanica (BBge) and Periplaneta americana (BPam), the sub social cockroach Cryptocercus punctulatus (BCpu) and the termite Mastotermes darwiniensis (BMda), show the remarkable stasis of this symbiotic system along the 140 Myr of evolutionary history of the cockroaches. The gene content as well as the chromosome architecture is highly maintained in the five studied genomes, being the most conserved among the known endosymbionts in which more than one strain have been analysed. Evolutionary tests point that purifying selection is the major evolutionary force operating in this system but few genes are accelerated. Stoichiometric analysis of the central metabolic pathways in the five strains of *Blattabacterium* shows a strict functional conservation in terms of biosynthetic precursors production, albeit some enzymatic steps were lost in the ancestors of BBor/BPam and BCpu/BMda, affecting the citric acid and the urea cycles, respectively. Our studies suggest that the genomic and metabolic architectures in *B. cuenoti* were sculpted very early after the establishment of its symbiosis with cockroaches.

Keywords: Blattabacterium pangenome, urease, urate metabolism

/	)
<u> </u>	 
1	
$\vdash$ $-$	 
1	
1	
1	
1	
<u> </u>	 
<u> </u>	 
$\vdash$ $-$	 
1	
1	
1	
$\vdash$ $-$	 
1	
1	
1	
1	
1	
1	
1	
1	
1	 i a
1	
1	
1	
1	
1	
1	
\	/
\	/
\	
	/

# The genome of *Candidatus Cardinium* hertigii, a secondary endosymbiont of the whitefly *Bemisia tabaci*

Santos-García D.¹, Beitia F.², Mouton L.³, Moya A.¹, Latorre A.¹and <u>Silva F.J.¹</u>

¹Institut Cavanilles de Biodiversitat i Biologia Evolutiva, University of Valencia, Spain; ²Instituto Valenciano de Investigaciones Agrarias (IVIA), Valencia, Spain; ³Laboratoire de Biométrie et Biologie Evolutive, UMR CNRS 5558, Université de Lyon, France - francisco.silva@uv.es

The whitefly Bemisia tabaci is a complex species composed by several biotypes, two of which, B and Q, have a wide distribution in Mediterranean countries. All individuals of this species present a gamma-proteobacterial P-endosymbiont, Portiera aleyrodidarum. Occasionally, the whiteflies can present also one or several S-symbionts from the genera Rickettsia, Hamiltonella, Wolbachia, Arsenophonus, Cardinium and Fritschea. While P. alevrodidarum is involved in supporting insect nutrition, the function of the S-symbionts needs to be characterized, although their different distribution among biotypes suggests that they might be contributing to different ecological traits. We have performed through a metagenomic approach the sequencing of the endosymbionts of a Q biotype strain which contains, with the obligate primary endosymbiont, two secondary endosymbionts: Hamiltonella defensa and Candidatus Cardinium hertigii. The genome of Candidatus Cardinium hertigii (endosymbiont of Bemisia tabaci) has been completely sequenced (average coverage 49). It is slightly higher than 1 Mb and, together with the chromosome, includes a circular plasmid of 53 kb. As expected for most endosymbionts the average G+C content of both is low (36%). The genome contains a large fraction of repeat sequences with more than 15% of the genome formed by transposable elements. The analysis of its gene repertoire gives clues about the relation with its whitefly host.

Keywords: symbiosis, Bemisia tabaci, Candidatus Cardinium hertigii, insect endosymbiont, genomics

/																													<hr/>
	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_			—	_	_	_	
-	—	_	—	—	—	—	—	—	—	—		—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	_		-
	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_
_	_	_	_	_	_	—	_	_	_	_	_	_	_	_	—	_	_	_	_	—	_	_	_	_	_	_	_	_	
-	—	_	_	—	—	—	_	_	—	—	_	—	_	—	—	_	—	_	—	—	_	—	—	—	—	—	_	_	-
	_	_	_	_	_	_	_	_	_		_	_	_		_	_	_	_		_	_	_	_			_	_	_	
-	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	-
	_	_	_	_	_	_	_	_	_			_	_	_	_	_	_	_	_	_	_	_	_		_	_	_	_	_
-	_	_	_	_	_	—	_	_	_		_	_	_	_	—	_	_	_	_	_	_	_	_		—	_	_	_	
-	_	_	_	_	—	_	_	_	_	_	_	_	_	—	—	_	_	_	—	—	_	_	_	_	—	_	_	_	_
	_	_	_	_	_		_	_	_	_	_	_	_	_		_	_	_	_	_	_	_	_	_	_	_	_	_	
F	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	-
$\vdash$	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	$\neg$
-	_					_	_	-	_	-	-	_	-	_	_	_	-	_	_	_	—	_	—	—	—		_	_	
-	_		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_			-,	_
																												A	
																											2		
																										//	1		
																										1	1		
																							~	~	~				
																							-						
/																													
																												/	·

# Distribution of *Wolbachia* strain wMelPop in *Drosophila* melanogaster brain cells analyzed with electron microscopy

Strunov A. and Kiseleva E.

### Russian Federation, 630090, Novosibirsk, Prospekt Lavrentyeva 10, The Institute of Cytology and Genetics, strunov.anton@gmail.com

Wolbachia are intracellular endosymbionts that can be found in a variety of host tissues including gonads, muscles and brain. Such a vast distribution of bacteria leads to a great diversity of *Wolbachia*-host interactions range from mutualism to parasitism and much depend on bacteria strain and species of the host. The wMelPop strain of *Wolbachia* is a striking example of pathogenic endosymbionts that over-replicates in host brain and muscle cells ultimately causing tissue degeneration and premature death. Understanding the features of *Wolbachia* negative impact on host organism is the research priorities due to the potential use of wMelPop strain as an environmentally friendly, bio-control agent.

The purpose of this study was to reveal the peculiarities of wMelPop morphology and its localization in *Drosophila* melanogaster brain cells with the advantages of transmission electron microscopy (TEM).

TEM analysis has shown that wMelPop is generally found in neuron cell body (soma) as well as in glial cells and sometimes in intercellular space but not presented in neuropil. According to the bacterial titer neurons can be divided into three types with different *Wolbachia* density per soma section. The bacterial distribution throughout the brain is very uneven and varies between individuals. In addition to the typical morphology the wMelPop strain shows different ultrastructural abnormalities like electron-dense matrix, bacterial outer membrane protrusion and enlargement as well as diverse vesicle inclusions. All these findings indicate that wMelPop is in an active state and has a predetermined pattern of distribution during the early embryogenesis in D. melanogaster brain cells.

#### Keywords: Wolbachia, Drosophila melanogaster, brain, TEM

This work was supported by the Program of the Presidium of RAS "Biodiversity" (26.30).

	/		
			1
	$\vdash$		
			1
		6	
·····			
			J
	\		/
	$\backslash$		/
			-

## Characterization of microbial community, associated with important forest pests (Lepidoptera)

Metla Z., Halimona J., Seskena R. and Jankevica L.

Laboratory of Experimental Entomology, Institute of Biology, University of Latvia, Miera iela 3, LV-2169 Salaspils, Latvia, e-mail: zmetla@inbox. lv

Laboratory of Experimental Entomology is working on problems, that include investigation of insect and entomopathogenic microorganism interactions to obtain a theoretical basis for effective development of microbiological methods for plant protection. Investigations are carried on of midgut microflora content and structure, their interactions and the effect on their changes, caused by environmental factors. Investigation of midgut microbial community and its diversity leads us to understanding the fundamental ecology of microorganisms in insect digestive tract. As test objects served pine looper (*Bupalus piniarius* L.), nun moth (Lymantria monacha L.), pine-tree lappet moth (Dendrolimus pini L.) and gypsy moth (*Lymantria dispar* L.).

Field-collected larvae from all Lepidoptera species were monitored for pathogen occurence in population and their role in it. All insects were screened by "culture dependant" method in order to acquire information of natural gut microflora content and identified entomopathogens. Morphologically different bacteria and fungi were isolated and classified by microscopy and physiological, biochemical reactions. The richness of microorganisms associated with fieldcollected larvae of Lepidoptera was identified. The results demonstrated that microbial flora of larvae is complex and dominanted by gammaproteobacteria and firmicutes class bacteria. Charecterization of gut microorganisms led us to identification of more than 100 different culturable isolates. The dominant bacteria species were Stenotrophomonas maltophilia, Serratia marcescens, Burkholderia cepacia, Corynebcaterium sp. and dominant fungi were Penicillium spp. All opportunistic entomopathogenic microorganism isolates from this study will be further evaluated for their suitability as potential virulent entomopathogens.

Keywords: Lepidoptera midgut, microbial diversity, entomopathogens

	/																													
1																														
	-	_	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	-
											_														_					
	-	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_		_	_	_	_	_
	-	_	_	_	_	—	_	_	_	_	—	—	_	_	_	—	_	_	_	_	—	_	_	_	—	—	—	_	_	_
	-		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	
			_	_	_		_	_	_			_	_	_		_	_	_	_		_	_	_	_		_	_	_	_	
	-	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_
-	-	_	—	—	—	—	—	—	_	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	_	_	-
	-	_	—	_	—	—	—	—	_	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	_	-
			_	_	_	_	-	_		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_			_	_	_	
1																														
		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_
$\vdash$	-	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	—	_	_	_	-
-	-	—	_	_			—	—	-	—	—	—	—	_	—	_	—	—	—	—	—	—	—	-	—	—	—	—	—	-
1																														
	-	_	_	_		_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_	-	-0	2
																												100		
																												~		
																												1		
																												1		
																										~!				
																								~	~					
																														1

#### A screen for host genes interacting with Wolbachia

Esteves S.S., Marialva M., Wilton J., Ferreira A.G. and Teixeira L.

#### Instituto Gulbenkian de Ciência, Oeiras, Portugal

Wolbachia are arguably the most widespread endosymbiotic bacteria known. Their presence can have profound effects on their hosts: from cytoplasmic incompatibility to protection from RNA viruses. However, little is known about the interaction of these bacteria with their hosts at the molecular level. We are approaching this problem taking advantage of the genetically tractable *Wolbachia*'s natural host *Drosophila* melanogaster. We are doing a genetic screen to identify host factors required by *Wolbachia* for a successful infection or required by the host to control or deal with this infection. We have screened approximately 700 lines and quantified the amount of *Wolbachia* in each of them. Interestingly, we do not observe a significant difference in *Wolbachia* titers between most of the different lines, suggesting that

*Wolbachia* levels are very well regulated. We are currently retesting some of the mutant lines where the *Wolbachia*-*Drosophila* tight regulation is disrupted.

Keywords: Wolbachia, Drosophila melanogaster

		<u> </u>
/		
(		
		1
		_
$\vdash$ $   -$		
	~~~~	
1		
\		/
\mathbf{i}		
		/

Enrichment strategy for sequencing of *Wolbachia* genomes from isopod crustaceans

Badawi M., Giraud I., Grève P. and Cordaux R.

Université de Poitiers

Laboratoire Ecologie et Biologie des Interactions - UMR CNRS 7267 Equipe Ecologie Evolution Symbiose - Bat. B8 - 40 avenue du Recteur Pineau - F-86022 POITIERS Cedex FRANCE e-mail : myriam.badawi@univ-poitiers.fr

It has been known for decades that Wolbachia endosymbionts induce reproductive manipulations such as feminization and cytoplasmic incompatibility (CI) in their isopod crustacean hosts. To date, the genetic basis for these manipulations remains unknown. To determine candidate genes, we plan to compare the genomes of two closely related Wolbachia strains infecting isopod species: the feminizing strain wVulC (host: Armadillidium vulgare) and the CI strain wCon (host: Cylisticus convexus). The acquisition of genome sequences requires purifying Wolbachia DNA, which currently remains a technical challenge. This is because Wolbachia bacteria are unculturable and extraction of total DNA results in high levels of contamination with nuclear and mitochondrial DNA from hosts. Indeed, our analysis by quantitative PCR of wCon samples revealed that for 1 base pair (bp) of Wolbachia sequence, there is typically ~300 bp of nuclear and mitochondrial sequence. To improve the ratio of Wolbachia to nuclear and mitochondrial sequence, we performed an artificial enrichment of the bacteria by shredding the cells without damaging nuclei and then separating host nuclei from bacteria by filtration. This resulted in an improved ratio of 1 bp of Wolbachia sequence for 1 bp of nuclear and mitochondrial sequence. Consistent results were obtained in replicates of several experiments. This ~300-fold enrichment makes our wCon sample amenable to 454 pyrosequencing. The next step will consist in mapping wCon reads against the previously sequenced wVulC genome to identify candidate genes. To conclude, this method of selective enrichment of Wolbachia DNA is efficient, fast, cheap, reliable, avoids centrifugation and DNase treatment, and can easily be extended to other Wolbachia strains of isopod crustaceans.

Keywords: Wolbachia ; Isopod ; DNA enrichment method; feminization, cytoplasmic incompatibility

		<u> </u>
/		
(
		1
		_
\vdash $ -$		
	~~~~	
1		
\		/
$\mathbf{i}$		
		/

### List of participants

Alex AEBI Agroscope Reckenholz-Tänikon Biosafety ZÜRICH - SWITZERLAND alexandre.aebi@art.admin.ch

Jaime AGUILERA European Food Safety Authority PARMA - ITALY jaime.aguilera@efsa.europa.eu

Serap AKSOY Yale School of Public Health New Haven, CT 06520-8034 serap.aksoy@yale.edu

Hisashi ANBUTSU National Institute of Advanced Industrial Science and Technology (AIST) TSUKUBA - JAPAN h-anbutsu@aist.go.jp

Sandra Breum ANDERSEN University of Copenhagen Centre for Social Evolution COPENHAGEN - DENMARK SBAndersen@bio.ku.dk

Telschow ARNDT Institute for Evolution and Biodiversity Genome Evolution MUENSTER - GERMANY a.telschow@wwu.de

Myriam BADAWI Université de Poitiers Laboratoire Ecologie et Biologie des Interaction - EBI - UMR 7267 POITIERS - FRANCE myriam.badawi@univ-poitiers.fr

Claudio BANDI Universita' Degli Studi Di Milano Dipartimento Di Patologia Animale Igiene E Sanita' Pubblica Veterinaria Sezione Di Parassitologia MILANO - ITALY claudio.bandi@unimi.it

Nicolas BECH Université de Poitiers Laboratoire Ecologie et Biologie des Interaction - EBI - UMR 7267 POITIERS - FRANCE nicolas.bech@univ-poitiers.fr Sophie BELTRAN Université de Poitiers Laboratoire Ecologie et Biologie des Interaction - EBI - UMR 7267 POITIERS - FRANCE sophie.beltran@univ-poitiers.fr

Joanne BERTAUX Université de Poitiers Laboratoire Ecologie et Biologie des Interaction - EBI - UMR 7267 POITIERS - FRANCE joanne.bertaux@univ-poitiers.fr

Karel BOLCKMANS Koppert B.V. BERKEL EN RODENRIJS - THE NETHERLANDS cteunissen@koppert.nl

Jacobus J. BOOMSMA Department of Biology, University of Copenhagen Centre for Social Evolution COPENHAGEN - DENMARK JJBoomsma@bio.ku.dk

Yuval BOAZ Hebrew University REHOVOT-ISRAEL yuval@agri.huji.ac.il

Didier BOUCHON Université de Poitiers Laboratoire Ecologie et Biologie des Interaction - EBI - UMR 7267 POITIERS - FRANCE didier.bouchon@univ-poitiers.fr

Kostas BOURTZIS University of Western Greece, Department of Environmental and Natural Resources Management AGRINIO - GREECE kbourtz@uoi.gr

Henk R BRAIG Bangor University School of Biological Sciences BANGOR - UK h.braig@bangor.ac.uk

Christine BRAQUART-VARNIER Université de Poitiers Laboratoire Ecologie et Biologie des Interaction - EBI - UMR 7267 POITIERS - FRANCE christine.braquart@univ-poitiers.fr Hans BREEUWER AMSTERDAM - THE NETHERLANDS j.a.j.breeuwer@uva.nl

Marie-Christine CARPENTIER Université Lyon 1 Laboratoire de Biométrie et Biologie Evolutive - UMR CNRS 5558 VILLEURBANNE - FRANCE marie-christine.carpentier@univ-lyon1.fr

Bodil CASS University of Arizona Departement of Entomologyr TUCSON, ARIZONA - USA bcass@email.arizona.edu

Yves CAUBET Université de Poitiers Laboratoire Ecologie et Biologie des Interaction - EBI - UMR 7267 POITIERS - FRANCE yves.caubet@univ-poitiers.fr

Ewa CHROSTEK Instituto Gulbenkian de Ciência Host-Microorganism Interactions OEIRAS - PORTUGAL echrostek@igc.gulbenkian.pt

Richard CORDAUX Université de Poitiers Laboratoire Ecologie et Biologie des Interaction - EBI - UMR 7267 POITIERS - FRANCE richard.cordaux@univ-poitiers.fr

Daniele DAFFONCHIO DISTAM, University of Milan MILAN - ITALY daniele.daffonchio@unimi.it

Alistair DARBY University of Liverpool Institute of Integrative Biology LIVERPOOL - UK acdarby@liv.ac.uk

Jessica DITTMER Université de Poitiers Laboratoire Ecologie et Biologie des Interaction - EBI - UMR 7267 POITIERS - FRANCE jessica.dittmer@univ-poitiers.fr Stephen DOBSON University of Kentucky LEXINGTON - USA sdobson@uky.edu

Sara EPIS Universita' Degli Studi Di Milano Dipartimento Di Patologia Animale Igiene E Sanita' Pubblica Veterinaria Sezione Di Parassitologia MILANO - ITALY sara.epis@guest.unimi.it

Sara ESTEVES Instituto Gulbenkian de Ciência Host-Microorganism Interactions OEIRAS - PORTUGAL sesteves@igc.gulbenkian.pt

Guido FAVIA University of Camerino Molecular Entomology Lab- School of Bioscience & Biotechnology CAMERINO - ITALY guido.favia@unicam.it

Heike FELDHAAR University of Bayreuth Animal Ecology I BAYREUTH - GERMANY feldhaar@uni-bayreuth.de

Heather FLORES Cornell University Aquadro/Barbash Labs ITHACA - USA haf22@cornell.edu

Horacio FRYDMAN Boston University Frydman Lab BOSTON - USA hfrydman@bu.edu

Takema FUKATSU National Institute of Advanced Industrial Science and Technology (AIST) Bioproduction Research Institute TSUKUBA - JAPAN t-fukatsu@aist.go.jp

Ingrid FAYE Department of Genetics, Microbiology and Toxicology Stockholms universitet STOCKHOLMS - SUEDE ingrid.faye@gmt.su.se Sandrine GENIEZ New England Biolabs/ LEES IPSWICH - USA geniez@neb.com

Lise-Marie GENTY Université de Poitiers Laboratoire Ecologie et Biologie des Interaction - EBI - UMR 7267 POITIERS - FRANCE lise.marie.genty@univ-poitiers.fr

Clément GILBERT Université de Poitiers Laboratoire Ecologie et Biologie des Interaction - EBI - UMR 7267 POITIERS - FRANCE clement.gilbert@univ-poitiers.fr

Pierre GRÈVE Université de Poitiers Laboratoire Ecologie et Biologie des Interaction - EBI - UMR 7267 POITIERS - FRANCE pierre.greve@univ-poitiers.fr

Tom GROOT Koppert B.V. BERKEL EN RODENRIJS - THE NETHERLANDS tgroot@koppert.nl

Roy GROSS University of Wuerzburg Chair of Microbiology WUERZBURG - GERMANY roy@biozentrum.uni-wuerzburg.de

Peter HAMMERSTEIN University Berlin Institute for Theoretical Biology, Humboldt BERLIN - GERMANY p.hammerstein@biologie.hu-berlin.de

Toshiyuki HARUMOTO National Institute of Advanced Industrial Science & Technology (AIST) Symbiotic Evolution & Biological Functions Research Group (Fukatsu Lab) TSUKUBA - JAPAN t-harumoto@aist.go.jp

Louise HEATON University of Sheffield Siva-Jothy Lab SHEFFIELD - UK I.heaton@sheffield.ac.uk Abdelaziz HEDDI INSA de Lyon UMR INRA/INSA de Lyon BF2I (Biologie Fonctionnelle, Insectes et Interactions) VILLEURBANNE - FRANCE Abdelaziz.Heddi@insa-lyon.fr

Helene HENRI Université Lyon 1 Laboratoire de Biométrie et Biologie Evolutive - UMR CNRS 5558 VILLEURBANNE - FRANCE helene.henri@univ-lyon1.fr

Jeremy HERREN Ecole Polytechnique Fédérale de Lausanne Lemaitre Lab LAUSANNE - SWITZERLAND jeremy.herren@epfl.ch

Matthias HORN University of Vienna Department of Microbial Ecology VIENNA - AUSTRIA horn@microbial-ecology.net

Greg HURST University of Liverpool Institute of Integrative Biology LIVERPOOL - UK g.hurst@liv.ac.uk

Filip HUSNIK University of South Bohemia Laboratory of molecular phylogeny and evolution of parasites CESKE BUDEJOVICE - CZECH REPUBLIC filip.husnik@gmail.com

Liga JANKEVICA Institute of Biology, University of Latvia Experimental entomology SALASPILS - LATVIA Jankevica.Liga@inbox.lv

Monique JOHNSON Université de Poitiers Laboratoire Ecologie et Biologie des Interaction - EBI - UMR 7267 POITIERS - FRANCE monique.johnson@univ-poitiers.fr

Edouard JURKEVITCH The Hebrew University of Jerusalem Microbial Ecology Lab REHOVOT - ISRAEL jurkevi@agri.huji.ac.il Lisa KLASSON Uppsala University Molecular evolution UPPSALA - SWEDEN lisa.klasson@icm.uu.se

Rok KOSTANJSEK Dept. of Biology, Biotechical faculty, University of Ljubljana LJUBLJANA - SLOVENIA rok.kostanjsek@bf.uni-lj.si

Wieslaw KOZEK Medical Sciences Campus, Univeristy of Puerto Rico SAN JUAN - UNITED STATES wjkozek@hotmail.com

Shoichiro KURATA Graduate School of Pharmaceutical Sciences, Tohoku University SENDAI - JAPAN kurata@mail.pharm.tohoku.ac.jp

Amparo LATORRE institute Cavanilles Of Biodiversity And Evolutionary Biology Evolutionary Genetics PATERNA, VALENCIA - SPAIN amparo.latorre@uv.es

Antonio LAZCANO Facultad de Ciencias, UNAM Origen de la Vida MEXICO D.F. - MEXICO alar@ciencias.unam.mx

Winka LE CLEC'H Université de Poitiers Laboratoire Ecologie et Biologie des Interaction - EBI - UMR 7267 POITIERS - FRANCE winka.leclech@yahoo.fr

Anders LIND Medical University of Vienna Lab Genome Dynamics VIENNA - AUSTRIA anders.lind@meduniwien.ac.at

Sandy MACDONALD YORK-USA sandyjmacdonald@gmail.com Thijs MACHTELINCKX Ghent University Agrozoology GHENT - BELGIUM thijs.machtelinckx@ugent.be

Isabelle MARCADE Université de Poitiers Laboratoire Ecologie et Biologie des Interaction - EBI - UMR 7267 POITIERS - FRANCE isabelle.marcade@univ-poitiers.fr

Julien MARTINEZ Department of Genetics, University of Cambridge Jiggins lab CAMBRIDGE - UNITED KINGDOM jtm35@cam.ac.uk

Humberto MARTINEZ Texas A&M University Mariana Mateos Lab COLLEGE STATION - U.S.A. lapostisima@tamu.edu

Patrick MAVINGUI CNRS Microbial Ecology - UMR 5557 VILLEURBANNE - FRANCE patrick.mavingui@univ-lyon1.fr

Adrien MERVILLE Université Lyon 1 Laboratoire de Biométrie et Biologie Evolutive - UMR CNRS 5558 VILLEURBANNE - FRANCE adrien.merville@univ-lyon1.fr

Zane METLA Institute of Biology, University of Latvia Laboratory of Experimental entomology SALASPILS - LATVIA zmetla@inbox.lv

Wolfgang MILLER Medical University of Vienna Lab Genome Dynamics VIENNA - AUSTRIA wolfgang.miller@meduniwien.ac.at

Guillaume MINARD CNRS-Université Lyon 1 UMR 5557 Ecologie Microbienne VILLEURBANES - FRANCE guillaume_minard_86@hotmail.com Yves MONE Université Lyon 1 Laboratoire de Biométrie et Biologie Evolutive - UMR CNRS 5558 VILLEURBANNE - FRANCE yves.mone@univ-lyon1.fr

Marta MOREIRA Universidade da Madeira ISOPlexis/Banco de Germoplasma FUNCHAL - PORTUGAL marta_amoreira@hotmail.com

Elmina MOTTIN Université de Poitiers Laboratoire Ecologie et Biologie des Interaction - EBI - UMR 7267 POITIERS - FRANCE elmina.mottin@univ-poitiers.fr

Bouziane MOUMEN Université de Poitiers Laboratoire Ecologie et Biologie des Interaction - EBI - UMR 7267 POITIERS - FRANCE bouziane.moumen@univ-poitiers.fr

Laurence MOUTON Université Lyon 1 Laboratoire de Biométrie et Biologie Evolutive - UMR CNRS 5558 VILLEURBANNE - FRANCE laurence.mouton@univ-lyon1.fr

Andrés MOYA Instituto Cavanilles de Biodiversidad y Biología Evolutiva, Universitat de València Evolutionary Genetics PATERNA, VALENCIA - SPAIN andres.moya@uv.es

Kaare NIELSEN University of Tromso Dept Pharmacy TROMSO - NORWAY kaare.nielsen@uit.no

Evangelos NTOUNTOUMIS University of Western Greece AGRINIO - GREECE edoudoum@cc.uoi.gr

Scott O'NEILL Monash University Scott O'Neill Lab MELBOURNE - AUSTRALIA scott.oneill@monash.edu Chrystel OLIVIER SASKATOON - CANADA chrystel.olivier@agr.gc.ca

Juan Camilo PAREDES Ecole Polytechnique Fédérale de Lausanne Lemaitre Lab LAUSANNE - SWITZERLAND juan.paredes@epfl.ch

JULI PERETO Institut Cavanilles de Biodiversitat i Biologia Evolutiva, Universitat de València Evolutionary Genetics PATERNA - SPAIN pereto@uv.es

M. Alejandra PEROTTI University of Reading School of Biological Sciences READING - UK m.a.perotti@reading.ac.uk

Jean POPOVICI Monash University O'Neill Lab MELBOURNE - AUSTRALIA jean.popovici@monash.edu

Maryline RAIMOND Université de Poitiers Laboratoire Ecologie et Biologie des Interaction - EBI - UMR 7267 POITIERS - FRANCE maryline.raimond@univ-poitiers.fr

Vincent RAQUIN CNRS - University Lyon 1 Microbial Ecology, Lyon VILLEURBANNE - FRANCE vincent.raquin@univ-lyon1.fr

Carolin RATZKA Universität Würzburg Lehrstuhl für Mikrobiologie WÜRZBURG - GERMANY carolin.ratzka@biozentrum.uni-wuerzburg.de

Freddie-Jeanne RICHARD Université de Poitiers Laboratoire Ecologie et Biologie des Interaction - EBI - UMR 7267 POITIERS - FRANCE freddie.jeanne.richard@univ-poitiers.fr Markus RIEGLER University of Western Sydney Hawkesbury Institute for the Environment PENRITH - AUSTRALIA m.riegler@uws.edu.au

Panagiotis SAPOUNTZIS CSE COPENHAGEN - DENMARK Sapountzis@bio.ku.dk

Davide SASSERA Università degli Studi di Milano DIPAV MILANO - ITALY davide.sassera@unimi.it

Sara SAVIC Scientific Veterinary Institute Novi Sad Laboratory for serology and immunology NOVI SAD - SERBIA sara@niv.ns.ac.rs

Daniela SCHNEIDER Medical University of Vienna Genome Dynamics VIENNA - AUSTRIA daniela.schneider@meduniwien.ac.at

Elena SHAIKEVICH Vavilov Institute of General Genetics Laboratory of Insect Genetics MOSCOW - RUSSIAN FEDERATION iaz34@mail.ru

Mathieu SICARD Université de Poitiers Laboratoire Ecologie et Biologie des Interaction - EBI - UMR 7267 POITIERS - FRANCE mathieu.sicard@univ-poitiers.fr

Francisco J. SILVA University of Valencia Evolutionary Genetics, Institut Cavanilles de Biodiversitat i Biologia Evolutiva PATERNA, VALENCIA - SPAIN francisco.silva@uv.es

Barton SLATKO New England Biolabs Molecular Parasitology IPSWICH MA - USA slatko@neb.com Catherine SOUTY-GROSSET Université de Poitiers Laboratoire Ecologie et Biologie des Interaction - EBI - UMR 7267 POITIERS - FRANCE catherine.souty@univ-poitiers.f

Anton STRUNOV Institute of Cytology and Genetics Laboratory of morphology and function of cell structures NOVOSIBIRSK - RUSSIAN FEDERATION strunov.anton@gmail.com

Luis TEIXEIRA Instituto Gulbenkian de Ciência Host-Microorganism Interactions OEIRAS - PORTUGAL Iteixeira@igc.gulbenkian.pt

Olle TERENIUS Swedish University of Agricultural Sciences Ecology UPPSALA - SWEDEN olle.terenius@slu.se

Gavin THOMAS University of York Department of Biology YORK - UK gavin.thomas@york.ac.uk

Amy TRUITT Portland State University De Rivera lab PORTLAND - UNITED STATES OF AMERICA amtruitt@pdx.edu

Claire VALIENTE MORO Université Claude Bernard Lyon 1 UMR CNRS 5557 Ecologie Microbienne VILLEURBANNE - FRANCE claire.valiente-moro@univ-lyon1.fr

Jan VAN DEN ABBEELE BELGIUM jvdabbeele@itg.be

Fabrice VAVRE Université Lyon 1 Laboratoire de Biométrie et Biologie Evolutive - UMR CNRS 5558 VILLEURBANNE - FRANCE fabrice.vavre@univ-lyon1.fr Christoph VORBURGER Institute of Integrative Biology, ETH Zürich & EAWAG DÜBENDORF - SWITZERLAND christoph.vorburger@eawag.ch

Jennifer WHITE University of Kentucky LEXINGTON, KY - UNITED STATES jenwhite.uk@gmail.com

Meg WOOLFIT Monash University Beth McGraw/Scott O'Neill MELBOURNE - AUSTRALIA meg.woolfit@monash.edu

Jason WULFF University of Kentucky, Department of Entomology White Lab LEXINGTON, KY - UNITED STATES jsnwlff@gmail.com

Ilya A. ZAKHAROV-GEZEKHUS Institute of General Genetics Animal Genetics MOSCOW - RUSSIA iaz34@mail.ru Einat ZCHORI-FEIN Agricultural Research Organization Entomology RAMAT YISHAY - ISRAEL einat@agri.gov.il

Flore ZÉLÉ CNRS Maladie infectieuse et vecteurs Ecologie Génétique Evolution Controle MONTPELLIER CEDEX 5 - FRANCE flore.zele@ird.fr

Renate ZINDEL Agroscope Reckenholz-Tänikon, ART Biosafety ZÜRICH - SWITZERLAND renate.zindel@art.admin.ch

Roman ZUG Institute for Theoretical Biology, Humboldt University Berlin BERLIN - GERMANY roman.zug@biologie.hu-berlin.de